

ISABELA BACALHAU DE OLIVEIRA

CRESCIMENTO E SOBREVIVÊNCIA LARVAL DO MARISCO
***Anomalocardia brasiliiana* (GMELIN, 1791) ALIMENTADO COM DIFERENTES**
DIETAS ALGAIS

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UNIVERSIDADE FEDERAL RURAL DE PERNAMBUCO
PRÓ-REITORIA DE PESQUISA E PÓS-GRADUAÇÃO
PROGRAMA DE PÓS-GRADUAÇÃO EM RECURSOS PESQUEIROS E AQUICULTURA

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***Anomalocardia brasiliiana* (GMELIN, 1791) ALIMENTADO COM DIFERENTES**
DIETAS ALGAIS

Isabela Bacalhau de Oliveira

Tese apresentada ao Programa de Pós-Graduação em Recursos Pesqueiros e Aquicultura da Universidade Federal Rural de Pernambuco como exigência para obtenção do título de Doutor.

Prof.(a) Dr.(a) Alfredo Olivera Gálvez
Orientador

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**CRESCIMENTO E SOBREVIVÊNCIA LARVAL DO MARISCO *Anomalocardia*
brasiliiana (GMELIN, 1791) ALIMENTADO COM DIFERENTES DIETAS ALGAIS**

Isabela Bacalhau de Oliveira

Tese julgada adequada para obtenção do título de doutor em Recursos Pesqueiros e Aquicultura. Defendida e aprovada em 25/08/2014 pela seguinte Banca Examinadora.

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Dedicatória

Dedico este trabalho as pessoas mais importantes de minha vida e que sempre estarão presentes em meu coração.

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Resumo

O presente estudo descreve a larvicultura do molusco bivalve *Anomalocardia brasiliiana*, com avaliação de diferentes dietas e densidade de estocagem. As dietas avaliadas foram: microalgas *Isochrysis galbana* (Ig), *Phaeodactylum tricornutum* (Phaeo), *Chaetoceros calcitrans* (Cca), *Pavlova lutheri* (Pl) e as combinações (Ig + Cca), (Ig + Phaeo), (Cca + Phaeo) e (Cca + Pl), totalizando oito dietas fornecidas por um período de 15 dias de cultivo. A microalga *I. galbana* fornecida isoladamente, apresentou menor sobrevivência e crescimento em relação as demais dietas testadas. O uso de *C. calcitrans* e *P. tricornutum* isoladamente ou combinada as outras microalgas apresentou melhores valores de sobrevivência e crescimento. A dieta combinada Cca + Pl, obteve maior crescimento e sobrevivência nas larvas de *A. brasiliiana* ($261,67 \pm 9,64 \mu\text{m}$ e $31,50 \pm 0,87 \%$). Todas as dietas avaliadas obtiveram resultados satisfatórios quanto ao crescimento e sobrevivência de larvas de *A. brasiliiana*, exceto a dieta *I. galbana* quando fornecida isoladamente. As densidades de estocagem avaliadas nas pós-larvas de *Anomalocardia brasiliiana* foi de 40, 80 e 160 ind.cm^{-2} em um período experimental de 28 dias. A densidade de 40 ind.cm^{-2} apresentou a maior taxa de crescimento específico diária. As maiores sobrevivências das pós-larvas foram observadas nas menores densidades $53,24 \pm 4,60 \%$ (40 ind.cm^{-2}) e $52,95 \pm 3,32 \%$ (80 ind.cm^{-2}), diferindo significativamente da maior densidade de estocagem com $31,54 \pm 0,70 \%$. Assim pôde-se concluir que podemos produzir larvas de *A. brasiliiana* utilizando qualquer uma das dietas avaliadas, exceto *I. galbana* se fornecida isoladamente. No cultivo de pós-larvas desta mesma espécie devemos realizar o manejo na densidade de estocagem no decorrer do crescimento, a densidade de 160 ind.cm^{-2} pode ser utilizada até que as larvas alcancem $600\mu\text{m}$ de comprimento, larvas maiores que $600 \mu\text{m}$ devem ser cultivadas na densidade de 40 ind.cm^{-2} para manter a taxa de crescimento máximo diária.

Palavras-chave: microalgas, densidade de estocagem, larvas, pós-larvas, manejo.

Abstract

The present study describes the larval rearing of bivalve mollusc *Anomalocardia brasiliiana*, with evaluation of different diets and stocking density. The diets were evaluated: microalgae *Isochrysis galbana* (Ig), *Phaeodactylum tricornutum* (Phaeo), *Chaetoceros calcitrans* (Cca), *Pavlova lutheri* (Pl) and combinations (Ig + Cca), (Ig + Phaeo) (Cca + Phaeo) and (Cca + Pl), totaling eight diets provided for a period of 15 days of cultivation. The microalgae *I. galbana* provided alone had lower survival and growth compared with other diets tested. The use of *C. calcitrans* and *P. tricornutum* alone or combined other microalgae showed highest values for survival and growth. The combined diet Cca + Pl, demonstrated the better growth and survival of larval *A. brasiliiana* ($261.67 \pm 9.64 \mu\text{m}$ and $31.50 \pm 0.87\%$). All diets evaluated with satisfactory results regarding the growth and survival of larvae of *A. brasiliiana* except *I. galbana* diet when given alone. The stocking densities evaluated in post-larvae *Anomalocardia brasiliiana* was 40, 80 and 160 ind.cm⁻² in a trial period of 28 days. The density of 40 ind.cm⁻² had the highest rate of daily specific growth. The highest survival of post-larvae were observed at lower densities $53.24 \pm 4.60\%$ (40 ind.cm⁻²) and $52.95 \pm 3.32\%$ (80 ind.cm⁻²), differing from greater stocking density with $31.54 \pm 0.70\%$. So we concluded that we can produce larvae of *A. brasiliiana* evaluated using any of the diets, except *I. galbana* was provided separately. In cultivation post-larvae of the same species must perform storage management in density during growth, density 160 ind.cm⁻² can be used to achieve the larvae 600 μm in length, greater than 600 μm larvae must be grown at a density of 40 ind.cm⁻² to keep the rate of maximum growth rate.

Key words: microalgae, stocking density, larvae, post-larvae, management.

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1 - Introdução

A produção aquícola mundial baseia-se no cultivo de seis grupos de organismos aquáticos: peixes de água doce, peixes diádromos, peixes marinhos, algas, crustáceos e moluscos. Em 2012 a produção mundial de moluscos foi de 15,2 milhões de toneladas, sendo mais que o dobro da produção de crustáceos com 6,4 milhões de toneladas (FAO, 2014).

No Brasil o cultivo de moluscos é representado por três espécies, a ostra japonesa (*Crassostrea gigas*) espécie exótica, a ostra nativa (*Crassostrea brasiliiana*) e a vieira (*Nodipecten nodosus*). Entretanto, existem várias outras espécies nativas que apresentam potencial para o cultivo dentre elas pode-se citar o marisco *Anomalocardia brasiliiana*, sendo cultivadas espécies da mesma família (Veneridae) em outras partes do mundo e que apresentam um mercado consolidado. O maior problema, para produção em laboratório, é conseguir uma produção regular de sementes, pois pouco se conhece sobre as exigências desta espécie para determinação de uma metodologia de produção.

As Microalgas marinhas são cultivadas para serem utilizadas como alimento para as várias fases de vida dos moluscos bivalves e, até o momento, têm constituído o principal alimento de larvas e sementes de bivalves cultivados (HELM et al., 2004). Outra função do cultivo de microalgas na aquicultura é proporcionar a melhoria da qualidade da água, através da absorção de produtos nitrogenados tóxicos (amônia e nitrito) e combate as bactérias patogênicas pela sua produção de substâncias antibióticas (LAVENS e SORGELLOS, 1996; REITAN et al. 1994). As microalgas mais utilizadas nas larviculturas de moluscos bivalves são as espécies dos seguintes gêneros: *Isochrysis*, *Pavlova*, *Chaetoceros*, *Phaeodactylum*, *Skeletonema*, *Thalassiosira*, *Dunaliella*, *Nannochloris*, *Tetraselmis* e *Rhodomonas* (COUTTEAU e SORGELOOS,

1992; MOUEZA et al., 1999; BROWN, 2002; HELM e BOURNE, 2004; CRAGG, 2006; LIU et al., 2009).

A larvicultura é uma das etapas mais importantes no cultivo de organismos aquáticos, e uma alimentação nutricionalmente balanceada, trará mais resistência aos animais quando forem cultivados fora do laboratório. No entanto, uma das principais causas de mortalidade na larvicultura é o desenvolvimento da comunidade bacteriana. Por exemplo larvas de vieira, *Pecten maximus*, são suscetíveis à contaminação bacteriana causando alta mortalidade nos cultivos intensos (COMELY, 1972; NICHOLAS et al., 1996). Além disso, fatores como alta temperatura, alta densidade de estocagem e excesso de nutrientes podem favorecer a proliferação de patógenos oportunistas (ANDERSEN et al., 2000).

O efeito da densidade de estocagem no crescimento e sobrevivência de larvas, juvenis e adultos de moluscos bivalves tem sido amplamente estudado (FRÉCHETTE, 2005), principalmente para aquelas espécies com importância comercial para aquicultura, como: *Crassostrea gigas*, *Ruditapes philippinarum*, *Perna perna*. A maioria dos estudos avaliaram o período de crescimento nas fases de D véliger até a metamorfose para pé de véliger (LIU et al., 2006; YAN et al., 2006; LIU et al., 2010), pois nesta fase as larvas são frágeis e altamente susceptíveis a doenças.

O conhecimento da melhor microalga, e/ou suas combinações na alimentação de *A. brasiliiana* assim como o manejo adequado é essencial para aumentar o desenvolvimento e sobrevivência de larvas e pós-larvas de marisco.

2 - Objetivos

2.1 - Objetivo geral

A presente proposta visa contribuir com o aprimoramento de tecnologias para a larvicultura do marisco *A. brasiliiana* na região Nordeste do Brasil.

2.2 - Objetivo(s) específico(s)

1. Avaliar o efeito de diferentes dietas microalgais no crescimento e sobrevivência de larvas do marisco *A. brasiliiana*;
2. Analisar o efeito de dietas combinadas de microalgas no crescimento e sobrevivência de pós-larvas do marisco *A. brasiliiana*
3. Avaliar o efeito da densidade de estocagem no crescimento e sobrevivência de pós-larvas do marisco *A. brasiliiana*;
4. Analisar o efeito da densidade de estocagem no crescimento e sobrevivência de pós-larvas do marisco *A. brasiliiana*, ao longo do tempo.

3 - Revisão de literatura

Anomalocardia brasiliiana pertence ao Reino Animal, Filo Mollusca, Classe Bivalvia, Subclasse Heterodonta, Infraclasse Euheterodonta, Ordem Veneroida, Superfamília Veneroidea, Família Veneridae, Gênero *Anomalocardia* e espécie *brasiliiana* (Gmelin, 1791). Reúnem aproximadamente 500 espécies viventes, pertencentes à aproximadamente cinquenta gêneros e doze subfamílias (CANAPA et al., 1996). Essa diversidade está relacionada à grande variedade de habitats que estão adaptados, tais como: praias arenosas, areno-lodosas, manguezais e fundos arenosos de ambientes coralíneos (CANTERA, 1991). No Brasil foram registradas 35 espécies de venerídeos, pertencentes a quatorze gêneros e sete subfamílias (RIOS, 1994).

A. brasiliiana vive em profundidades de 0,3 – 5 m, superficialmente escavado na areia próximo ao manguezal, sensíveis a variações ecológicas, com alta mortalidade devido às chuvas, causando grandes flutuações no tamanho e na distribuição das populações (MOUEZA et al., 1999; MONTI et al., 1991). Distribui-se desde a costa das Antilhas até o Uruguai (RIOS, 1994), e o tamanho máximo já encontrado foi de 38 mm segundo dados do Malacolog version 4.1.1. (A Database of Western Atlantic Marine Mollusca). Encontra-se em maior abundância na região entre marés nos bancos naturais formados da interação dos mesmos no sedimento com uma grande variedade de bancos naturais, *micro-habitats*. É conhecido popularmente na região sudeste e sul do Brasil como berbigão, papafumo, sarnambi ou vôngoli, e na região nordeste é chamado de marisco-pedra e maçunim (OLIVEIRA et al., 2014).

A *A. brasiliiana* é uma espécie dióica, ou seja, apresenta sexos separados, não apresenta características morfológicas externas (diferença na coloração ou tamanho das conchas) ou internas (diferença na coloração das gônadas) que permitam a diferenciação macroscópica dos sexos, sendo necessária análise microscópica dos gametas ou estudos

histológicos (GROTTA & LUNETTA, 1980). A Reprodução é sexuada com lançamento de gametas na água e fecundação externa.

O desenvolvimento larval da *A. brasiliiana* passa pelas fases trocófora, véliger (D-véliger, umbonada e pé-de-véliger) e pós-larva ou juvenil. MOÛEZA et al (1999) relataram o surgimento de larvas trocóforas após 9 horas do início da fertilização, e após 24 h de vida já apresentavam aproximadamente 95 µm de comprimento e estavam no estágio D-véliger. A partir do quinto dia passaram para véliger com comprimento médio de 150 µm. No sétimo dia inicia a metamorfose para fase pé de véliger, sendo esta fase considerada crítica, pois as larvas são frágeis e susceptíveis a doenças durante o manejo (OLIVEIRA et al., 2007). Aos 11 dias de vida, as larvas migram para o fundo, fase pé de véliger, 180 µm de comprimento, e com 15 dias apresentando todas as características de juvenil, sifão e o pé totalmente desenvolvidos, com aproximadamente 300 µm de comprimento (MOÛEZA et al., 1999).

No habitat natural a espécie pode reproduzir o ano todo, porém apresenta picos de reprodução em algumas épocas do ano. O período reprodutivo para uma mesma espécie pode variar substancialmente de região para região, principalmente em função de diferentes condições climáticas e ambientais (LAVANDER et al., 2011). Sendo o Brasil um país com grande extensão costeira, possivelmente o padrão maturacional para *A. brasiliiana* é diferenciado entre as regiões do país.

Estudos realizados sobre a biologia reprodutiva da *A. brasiliiana* no Brasil identificaram que em São Paulo, Santa Catarina e no Paraná a espécie apresenta ciclo reprodutivo contínuo, com dois períodos de liberação de gametas na primavera e outono, assim como um período de repouso parcial no inverno (NARCHI, 1976; BOEHS, 2000; ARAÚJO, 2001).

No estado da Paraíba, a produção de gametas ocorre durante todo o ano, devido às condições ambientais favoráveis, temperatura constante e baixo índice de precipitação (GROTTA & LUNETTA, 1980). No Ceará foi observado dois picos reprodutivos: julho a outubro (primavera e inverno) e fevereiro a abril (verão e outono) (BARREIRA & ARAÚJO, 2005). Em Pernambuco, da mesma forma, apresenta ciclo reprodutivo contínuo, com eliminação de gametas de outubro a junho (LAVANDER et al., 2011).

Concomitantemente aos estudos de biologia reprodutiva foram realizados estudos sobre a dinâmica populacional de *A. brasiliiana* na costa brasileira (PEZZUTO e ECHTERNACHT, 1999; BOEHS, 2008; RODRIGUES, 2009; EL-DEIR, 2009; OLIVEIRA et al., 2011; RODRIGUES et al., 2013). Em Pernambuco, El-deir (2009) observou grandes variações na distribuição da espécie, apresentando uma densidade média de 1348 ind.m². Para o mesmo Estado, Oliveira et al. (2011) encontraram valores médios de densidade no período de verão e inverno de 298 ind.m² e 173 ind.m² respectivamente. Esses resultados são semelhantes aos encontrados por Rodrigues et al. (2013), que também estudaram a espécie no litoral de Pernambuco e verificou que em ambiente natural a espécie pode ser encontrada em até 268 ind.m².

Os dados de dinâmica populacional e biologia reprodutiva são necessários para o estabelecimento de medidas regulatórias da atividade pesqueira. Contudo, até o momento, as medidas legais tomadas não têm apresentado um resultado satisfatório na preservação dos estoques naturais da *A. brasiliiana*.

A crescente demanda por frutos do mar e a facilidade com que os estoques costeiros podem ser capturados (DAME et al., 2002) resultou em uma super exploração e colapso de algumas espécies de pesca dos ecossistemas costeiros (MORA et al., 2009; FAO, 2010). Na costa brasileira, a pesca de *A. brasiliiana* é amplamente difundida e intensa, com grande importância econômica para grupos de famílias de pescadores

artesanais (OLIVEIRA et al., 2014). A coleta de *A. brasiliiana* está registrada desde 1970 em Pernambuco, quando a atividade era praticada apenas por mulheres, porém ao longo dos anos, o número de pescadores do sexo masculino tem aumentado significativamente (SILVA-CAVALCANTI, 2009).

Dentre as principais espécies de moluscos, no estado de Pernambuco, o marisco *A. brasiliiana* é o que mais se destaca. A produção no ano de 2007 foi de 2.479,2 t responsável por 20% da produção total de moluscos (CEPENE, 2008). É o principal recurso pesqueiro entre os bivalves, e a principal fonte de renda de marisqueiras, e teve sua exploração quase que dobrada entre os anos de 2003 a 2005 (PEZZUTO & ECHTERNACHT, 1999). Infelizmente não há dados recentes sobre a produção pesqueira de *A. brasilaiana* no estado de Pernambuco, porém as comunidades tradicionais perceberam a diminuição deste recurso tanto em relação ao tamanho quanto ao volume (SILVA-CAVALCANTI, 2009). ARRUDA-SOARES et al. (1982) recomendam a captura de espécimes de *A. brasiliiana* com comprimento acima de 20 mm, neste tamanho os indivíduos já teriam alcançado um grau de desenvolvimento reprodutivo que possibilitasse a reposição e manutenção dos estoques naturais.

O declínio dos bancos naturais de moluscos bivalves por todo mundo, tornou-se um insentivo ao crescimento das larviculturas para o fornecimento de sementes (PRADO et al., 2010). As atividades básicas de qualquer larvicultura de moluscos bivalves são condicionamento e desova do estoque reprodutor, cultivo e assentamento das larvas e sementes até um tamanho aceitável, assim como a produção de grandes quantidades de alimento (microalgas) para todas as fases do ciclo de produção (HELM et al., 2004).

A microalga *Isochrysis galbana* é considerada um ótimo alimento para as larvas dos moluscos bivalves *Mercenaria mercenaria* e *Tapes semidecussata*, porém para as

ostras *Crassostrea gigas* e *Crassostrea rhizophorae* e o marisco *Donax trunculus* não foi obtido um bom desenvolvimento larval (HELM e LAING, 1987; RUIZ-AZCONA et al., 1996). Liu et al. (2009) avaliaram o efeito de dietas de microalgas no crescimento e sobrevivência de larvas e pós-larvas de *Clinocardium nuttallii*. Os autores constataram que o uso de dietas espécie-específica de *I. galbana* (TISO) e *Chaetoceros muelleri* (CM) são suficientes para proporcionar um máximo crescimento de larvas e pós-larvas de *C. nuttallii*. Entretanto o uso destas microalgas utilizadas de forma combinada apresentou um melhor crescimento na fase larval ou no início da fase pós-larval, sendo recomendado o uso desta dieta bialgal para atender as necessidades nutricionais das sementes de *C. nuttallii*.

Martínez-Fernandez e Southgate (2007) obtiveram uma maior taxa de crescimento na fase D-larva de *Pinctada margaritifera* para aquelas cultivadas com a combinação ternária de *Pavlova* sp. / *Pavlova salina* / TISO. No entanto, esta taxa de crescimento não foi significativamente maior do que a das larvas alimentadas com a combinação binária do *Pav. salina* / TISO que tem sido usado como uma “dieta padrão” para a larvicultura de *P. margaritifera* (SOUTHGATE e BEER, 1997). Martínez-Fernandez e Southgate (2007) sugerem que a alimentação com uma única espécie de microalga para larvas de *P. margaritifera* pode ter uma finalidade prática, durante os primeiros 10 dias de larvicultura. No entanto, a adição de uma microalga diatomácea flagelada na dieta teve um incremento na taxa de crescimento e sobrevivência das larvas umbonadas de *P. margaritifera* quando comparada às combinações sem diatomáceas.

As haptophyta da classe Pavlovophyceae são microalgas flageladas com tamanho de 3-5 µm, ou seja apropriado para ingestão de larvas de moluscos (PONIS et al., 2006), são ricas em ácidos graxos polinsaturados (PUFA) (KANAZAWA et al., 1979; VOLKMAN et al., 1991). *Pavlova lutheri* é conhecida como um excelente alimento de

larvas de *Pecten maximus* e *Pecten fumatus*, contudo para larvas de *Crassostrea gigas* apresentou baixo valor alimentar, com menor sobrevivência das larvas (HEASMAN et al., 2000; PONIS et al., 2003a,b; LAING, 2004).

A utilização de *Pavlova pinguis* e *Radomonas salina* como alimento complementar a alimentação natural de juvenis da ostra do pacífico, *Crassostrea gigas*, em uma larvicultura comercial, produz aumento de custo-benefício na produção animal (BROWN et al., 1998). Estes mesmos autores observaram que a utilização de dietas mono-específica de *Tetraselmis* sp. CS-362 e *Nannochloropsis* sp. CS-246 foram mal ingeridas, e é provável que este fator tenha causado crescimento inferior das ostras.

Rivero-Rodríguez et al. (2007) constataram que a dieta mono-específica de *Chaetoceros calcitrans* para juvenis de *Crassostrea corteziensis* foi superior a outras dietas testadas (*Tetraselmis suecica*, *Isochrysis galbana* clone T-iso, *Phaeodactylum tricornutum*, *Chaetoceros muelleri*, *Chaetoceros calcitrans* e suas combinações bi-algaes). No geral, os juvenis de *C. corteziensis* alimentados com as dietas com qualquer combinação de *C. calcitrans* obtiveram melhor crescimento. O contrário foi observado para as microalgas *I. galbana*, *T. suecica*, *P. tricornutum* e suas combinações bi-algais.

Estudos afirmam que o uso da microalga *P. tricornutum*, em outros bivalves, afeta o crescimento, deixando-o mais lento (EPIFANIO et al., 1981; ALBENTOSA et al., 1996; RIVERO-RODRIGUEZ et al., 2007), devido à microalga ser de difícil digestão (RIVERO-RODRIGUEZ et al., 2007), possivelmente devido a falta de triptofano na dieta (EPIFANIO et al., 1981). Tang et al. (2006) obtiveram baixo crescimento de larvas de *Meretrix meretrix* quando alimentadas com *P. tricornutum* e *Pavlova viridis*.

O sistema Europeu de cultivo berçário de larvas de vieiras, inclui a troca completa da água várias vezes durante a semana, e alimentação uma ou duas vezes ao dia

(BUESTEL et al., 1982; COCHARD e DEVAUCHELLE, 1993; TAYLOR et al., 1994), que resulta em altos valores de matéria orgânica, e um ambiente microbiano instável, o qual é comumente controlado com uso profilático de agentes antibacteriano (ROBERT et al., 1996; TORKILDSEN et al., 2000). As larviculturas de moluscos bivalves são frequentemente afetadas por surtos de doenças, como infecções bacterianas do gênero *Vibrio* (PRADO et al, 2010). Andersen et al. (2000) observaram que o uso de antibióticos pode, hipoteticamente, alterar a atividade bacteriana ou a composição das espécies no sistema, ocorrendo menor degradação ou aumento na produção de amônia.

Prado et al. (2010) observaram em sua revisão que todos os tratamentos considerados (Filtração, Radiação Ultravioleta e Quimioterápicos) tem como objetivo a eliminação completa da microbiota presente na água, entretanto populações de bactérias ou parte delas tem efeito benéfico no desenvolvimento larval de bivalves. As bactérias parecem satisfazer as exigências metabólicas, fornecendo vitaminas ou outros fatores de crescimento (PRIEUR et al., 1990).

Foram realizados diversos estudos para estabelecer um protocolo de produção de larvas de bivalves, incluindo bivalves de areia (LIU et al, 2002; LIU et al, 2006; YAN et al, 2006; LIU et al, 2008; LIU et al, 2010), mexilhões (GALLEY et al, 2010; PETTERSEN et al, 2010), ostras (RICO-VILLA et al, 2006) e vieiras (RUPP e PARSONS, 2004; GOUDA et al, 2006), pois a identificação dos fatores que influenciam no crescimento e na sobrevivência das larvas de moluscos é essencial para o sucesso da larvicultura.

Fatores como temperatura, salinidade e alimentação podem influenciar na sobrevivência, crescimento e período de metamorfose larval para assentamento na espécie *A. brasiliana*. A sobrevivência e crescimento de moluscos são afetados por parâmetros físicos como temperatura e salinidade (KINNE, 1964), assim como manejo

e alimentação (DEVAKIE & ALI, 2000; OLIVEIRA et al, 2007). Para o molusco de areia mais cultivado no mundo, *Ruditapes philippinarum*, Delgado & Camacho (2007) descreveram que o aumento da temperatura influencia grande parte dos processos fisiológicos, afetando seu crescimento. Em *A. brasiliana* a temperatura também é fator primordial para seu crescimento (BARREIRA & ARAUJO, 2005).

Outra importante variável a ser considerada, é a densidade de estocagem em larvas de moluscos bivalves. O efeito da densidade no crescimento e sobrevivência de larvas, juvenis e adultos tem sido amplamente estudado (FRÉCHETTE, 2005), principalmente nas espécies com importância comercial para aquicultura. A maioria dos estudos avaliaram o período de crescimento nas fases de D véliger até a metamorfose para pé de véliger (LIU et al., 2006; YAN et al., 2006; LIU et al., 2010), pois é considerada crítica, devido as larvas serem frágeis e susceptíveis a doenças durante o manejo.

Yan et al., (2006) avaliaram a dieta, densidade de estocagem, intensidade luminosa, filtração da água, troca de água e sedimento, tendo obtido que a densidade de 5-10 larvas.mL⁻¹ mantém um crescimento normal em larvas de *Ruditapes philippinarum*. Para larvas de *Meretrix meretrix* a densidade de estocagem recomendada é de 10-20 larvas.mL⁻¹ (LIU et al., 2006), enquanto que para larvas de *Clinocardium nuttallii* a densidade de estocagem deve ser igual ou inferior a 4 larval.mL⁻¹ e ao se aproximar da metamorfose para a pé de véliger, reduz a densidade para 2,5 larvas.mL⁻¹ (LIU et al., 2010).

Quando as larvas de moluscos bivalves realizam a metamorfose para a etapa pé de véliger, estas tornam-se mais resistentes ao manejo e passam a ser cultivadas em densidades mais elevadas, por um período maior de tempo (semanas) até alcançarem o tamanho de semente (1mm) quando poderão ser cultivadas a campo (NICOLAS e

ROBERT, 2001; EPELBAUM et al., 2011; LIU et al., 2011). Portanto avaliar a densidade de estocagem durante a fase pós-larval, tem como finalidade promover um ambiente de cultivo adequado para que estas alcancem a fase de semente em um menor tempo possível.

Para larvas de *Clinocardium nuttallii* é recomendado a densidade de estocagem de 160 ind.cm⁻² até alcançarem 1 mm de comprimento, passando para 40 ind.cm⁻² até 2 mm, depois 20 ind.cm⁻² até 3 mm, finalizando com 10 ind.cm⁻² até 4 mm (LIU et al., 2011). Resultados similares foram encontrados por Jones et al. (1993), sendo 150-200 pé de veliger.cm⁻² de Manila clam *Tapes philippinarum* em down-wellers.

Uma larvicultura comercial tem como meta melhorar o cultivo, aumentando a produção por unidade em um volume limitado de água com o menor custo possível. Nas larviculturas de moluscos é comum utilizar os sistemas de fluxo contínuo de água, diferenciados pelo sentido da corrente, sentido de cima para baixo (Down-welling) e de baixo para cima (Up-welling). Contudo quando se utiliza unidades de pequeno volume, o sistema de cultivo fechado com circulação da água também é bastante utilizado.

4. - Referências bibliográficas

ALBENTOSA, M., PÉREZ-CAMACHO, A., LABARTA, V., FERNÁNDEZ-REIRIZ, M.J. Evaluation of live microalgal diets for the seed culture of *Ruditapes decussatus* using physiological and biochemical parameters. **Aquaculture**. (148): 11-23. 1996.

ANDERSEN, S.; BURNELL, G.; BERGH, Ø. Flow-through systems for culturing great scallop larvae. *Aquaculture International*. 8. 249-257. 2000.

ARAÚJO, C. M. Biologia reprodutiva do berbigão *Anomalocardia brasiliiana* (Mollusca: Bivalvia, Veneridae) na Reserva Extrativista Marinha do Pirajubaé (REMAPI) Estado de Santa Catarina. Florianópolis. 2001. 203p. (**Tese de Doutorado**). Universidade de São Paulo. São Paulo.

ARRUDA-SOARES, H.; SCHAEFFER-NOVELLI, Y. & MANDELLI JUNIOR, J. “Berbigão” *Anomalocardia brasiliiana* (Gmelin, 1791), bivalve comestível da região da Ilha do Cardoso, Estado de São Paulo, Brasil: Aspectos biológicos de interesse para a pesca comercial. **Boletim do Instituto Pesca**. v. 9, p. 21-38, 1982.

BARREIRA, C. A. R.; ARAÚJO, M. L. R. Ciclo reprodutivo de *Anomalocardia brasiliiana* (Gmelin, 1791) (Mollusca, Bivalvia, Veneridae) na Praia do Canto da Barra, Fortim, Ceará, Brasil. **Boletim do Instituto de Pesca**, São Paulo, 31(1): 9-20, 2005.

BOEHS, G. Ecologia populacional, reprodução e contribuição em biomassa de *Anomalocardia brasiliiana* (Gmelin, 1791) (Bivalve: Veneridae) na Baía de Paranaguá, Paraná, Brasil. 2000. 201 p. (**Tese de Doutorado**), Programa de Pós- Graduação em zoologia, Universidade Federal do Paraná, Curitiba.

BOEHS, G.; ABSHER, T. M. & CRUZ-KALED, A. C. Ecologia Populacional de *Anomalocardia brasiliiana* (Gmelin, 1791) (Bivalvia, Veneridae) na Baía de Paranaguá, Paraná, Brasil. **Boletim Instituto de Pesca**, São Paulo, 34(2): 259 – 270. 2008.

BUESTEL, D.; COHARD, J.-C.; DAO, J.-C.; GÉRARD, A. Production artificielle de naissain de coquilles Saint-Jacques *Pecten maximus* (L.). Premiers resultants en rade de Brest, Vie Marine. 4. 24-28. 1982. In: ANDERSEN, S.; BURNELL, G.; BERGH, Ø. Flow-through systems for culturing great scallop larvae. **Aquaculture International**. 8. 249-257. 2000.

BROWN, M.R. Nutritional value and use of microalgae en aquaculture. 2002. In: Cruz-Suárez, L. E., Ricque-Marie, D., Tapia-Salazar, M., Gaxiola-Cortés, M. G., Simoes, N. (Eds.). **Avances en Nutrición Acuicola VI**. Memorias del VI Simposium Internacional de Nutrición Acuicola. 3 al 6 de Septiembre del 2002. Cancún, Quintana Roo, México.

BROWN, M.R.; MCCAUSLAND, M.A.; KOWALSKI, K. The nutritional value of four Australian microalgal strains fed to Pacific oyster *Crassostrea gigas* spat. **Aquaculture**. 165, 281–293. 1998.

CANAPA, A.; MAROTA, I.; ROLLO, F. & OLMO, E. Phylogenetic analysis of Veneridae (Bivalvia): comparison of molecular and paleontological data. **J. Mol. Evol.** 43: 517-522. 1996.

CANTERA, J.R. Shallow-water venerid clams (Bivalvia: Veneridae) from the pacific coast of Colombia. *The Veliger*. 34:78-84. 1991.

CEPENE. - Boletim estatístico da pesca marítima e estuarina do Nordeste do Brasil - 2006. Tamandaré: Centro de Pesquisa e Gestão de Recursos Pesqueiros do Litoral Nordeste (CEPENE) 385p. 2008.

COCHARD, J.C.; DEVAUCHELLE, N. Spawning, fecundity and larval survival and growth in relation to controlled conditioning in native and transplanted populations of *Pecten maximus* (L.); evidence for the existence of separate stocks. **Journal of Experimental Marine Biology and Ecology**. 169. 41-56. 1993.

COUTTEAU, P.; SORGELOOS, P. The requirement for live algae and their replacement by artificial diets in the hatchery and nursery rearing of bivalve molluscs: an international survey. **Journal Shellfish Research**. 11. 467-476. 1992.

DAME, R.; BUSHEK, D.; ALLEN, D.; LEWITUS, A.; EDWARDS, D.; KOEPFLER, E.; GREGORY, L. Ecosystem response to bivalve density reduction: management implications. **Aquatic Ecology**, v.36, n.1, p.51-65, 2002.

CRAGG, S.M. Development, physiology, behaviour and ecology of scallop larvae. In: Shumway, S.E., Parsons, G.J. (Eds.), **Scallops: Biology, Ecology and Aquaculture**. Elsevier, B.V. Amsterdam. 102 pp. 2006.

GROTTA, M. & LUNETTA, J.E. Ciclo sexual de *Anomalocardia brasiliiana* (Gmelin, 1791) do litoral do Estado da Paraíba. **Rev. Nordest. Biol.** 3(1): 5-55. 1980.

DELGADO, M.; CAMACHO, A.P. Influence of temperature on gonadal development of *Ruditapes philippinarum* (Adams and Reeve, 1850) with special reference to ingested food and energy balance. **Aquaculture**. (264): 398-407. 2007.

DEVAKIE, M.; A. ALI. Effects of storage temperature and duration on the setting and post-set spat survival of the tropical oyster, *Crassostrea iredalei*. **Aquaculture**. (190): 369–376. 2000

EL-DEIR, S.G. Estudo da mariscagem de *Anomalocardia brasiliiana* (Mollusca Bivalvia) nos bancos de Coroa do Avião, Ramalho e Mangue Seco (Igarassu, Pernambuco, Brasil). 2009.123 p. **Tese de Doutorado**, Programa de Pós graduação em Oceanografia, Universidade Federal de Pernambuco, Recife.

EPIFANIO, C.E., VALENTI, C.C., TURK, C.L. A comparison of *Phaeodactylum tricornutum* and *Thalassiosira pseudonana* as foods for the oyster *Crassostrea virginica*. **Aquaculture**. (23): 247–253. 1981.

EPELBAUM, A., PEARCE, C.M., YUAN, S., PLAMONDON, N., GURNEY-SMITH, H. Effects of stocking density and substratum on the survival, growth, burrowing behaviour and shell morphology of juvenile basket cockle, *Clinocardium nuttallii*: implications for nursery seed production and field out planting. **Aquac. Res.** 42, 975-986. 2011.

FAO. **The State of World Fisheries and Aquaculture**. Rome. Food and Agriculture Organization of The United Nations. Rome. 218p. 2010.

FAO. **The state of world fisheries and aquaculture**. Food and Agriculture Organization of the United Nations. Rome. 243 p. 2014.

FRÉCHETTE, M. A comment on the methodology of stocking experiments. **Aquaculture**. 250, 291–299. 2005.

GALLEY, T.H., BATISTA, F.M., BRAITHWAITE, R., KING, J., BEAUMONT, A.R. Optimisation of larval culture of the mussel *Mytilus edulis* (L.). **Aquaculture International**. (18):315–325. 2010.

GOUDA, R., KENCHINGTON, E., HATCHER, B., VERCAEMER, B. Effects of locally-isolated micro-phytoplankton diets on growth and survival of sea scallop (*Placopecten magellanicus*) larvae. **Aquaculture**. 259, 169–180. 2006.

HELM, M.M., BOURNE, N. Hatchery culture of bivalve: a practical manual. FAO. **Fisheries Technical Paper**. 471, Rome, 177 p. 2004.

HELM, M.M.; LAING, I. Preliminary observations on the nutritional value of “Tahiti *Isochrysis*” to bivalve larvae. **Aquaculture**. 62. 281-288. 1987.

HEASMAN M.P., DIEMAR, J., O’CONNOR, W., SUSHAMES, T., FOULKE, L. Development of extended shelf-life microalgae concentrate diets harvested by centrifugation for bivalve mollusks: a summary. **Aquaculture Research**. (31):6, 37–59. 2000.

JONES, G.G., SANFORD, C.L., JONES, B.L. **Manila Clams: Hatchery and Nursery Methods**. Innovative Aquaculture Products Ltd. Skerry Bay. Lasqueti Island BC V0R 2J0. Canada. 73 pp. 1993.

KANAZAWA, A., TESHIMA, S., ONO, K. Relationships between the essential fatty acid requirement of aquatic animals and their capacity for bioconversion of linoleic acid to highly unsaturated fatty acid. **Comp. Biochem. Physiol.** 63B, 295–298. 1979.

KINNE, O. The effects of temperature and salinity on marine and brackish water animals. II. Salinity and temperature salinity combinations. **Oceanography Marine Biology**. Annual Ver. 2: 53 – 55. 1964.

LAING, I. Filtration of king scallops (*Pecten maximus*). **Aquaculture**. 240, 369–384. 2004.

LAVANDER, H. D.; CARDOSO JÚNIOR, L. O.; OLIVEIRA, R. L.; DA SILVA NETO, S. R.; GALVEZ; A. O.; PEIXOTO. S. R. M. Biologia reprodutiva da *Anomalocardia brasiliiana* (Gmelin, 1791) no litoral norte de Pernambuco, Brasil. **Revista Brasileira de Ciências Agrárias**, Recife, v.6, n.2, p. 344-350, 2011.

LAVENS, P., SORGELLOOS, P. **Manual on the production and use of live food for aquaculture**. FAO Fisheries Technical paper. In: Lavens P, Sorgeloos P (eds) Rome. 36–19 pp. 1996.

LIU, B., DONG, B., TANG, B., ZHANG, T., XIANG, J. Effect of stocking density on growth, settlement and survival of clam larvae, *Meretrix meretrix*. **Aquaculture** 258, 344–349. 2006.

LIU, W., MA, Y., HU, S., MIAO, G., LI, J. Rearing Venus Clam seeds, *Cyclina sinensis* (Gmelin), on a commercial scale. **Aquaculture** 211, 109-114. 2002.

LIU, W., ALABI, A.O., PEARCE, C.M. Broodstock conditioning in the basket cockle *Clinocardium nuttallii*. **J. Shellfish Res.** 27, 399–404. 2008.

LIU, W.; PEARCE, C.M.; ALABI, A.O.; GURNEY-SMITH, H. Effects of microalgal diets on the growth and survive of larvae and pos-larvae of the basket cockle, *Clinocardium nuttallii*. **Aquaculture**. 293. 248-254. 2009.

LIU, W.; GURNEY-SMITH, H.; BEERENS, A.; PEARCE, C.M. Effects of stocking density, algal density, and temperature on growth and survival of larval of the basket cockle, *Clinocardium nuttallii*. **Aquaculture**. 299. 99-105. 2010.

LIU, W., PEARCE, C.M., ALABI, A.O., BEERENS, A., GURNEY-SMITH, H.. Effects of stocking density, ration, and temperature on growth of early post-settled juveniles of the basket cockle, *Clinocardium nuttallii*. **Aquaculture**. 320, 129–136. 2011.

MARTÍNEZ-FERNÁNDEZ, E.; SOUTHGATE, P.C. Use of tropical microalgae as food for larvae of the black-lip pearl oyster *Pinctada margaritifera*. **Aquaculture**. 263. 220–226. 2007.

MONTI, D.; FRENKIEL, L.; MOUEZA, M. Demography and growth of *Anomalocardia brasiliiana* (Gmelin) (Bivalvia: Veneridae) in a Mangrove, in Guadeloupe (French West Indies). **Journal of Molluscs Studies**. 57, 249-257. 1991.

MORA, C.; MYERS, R. A.; COLL, M.; LIBRALATO, S.; PITCHER, T. J.; SUMAILA, R. U.; ZELLER, D.; WATSON, R.; GASTON, K. J.; WORM, B. Management effectiveness of the World's Marine Fisheries. **PLoS Biol** v.7, n.6, e1000131. 2009.

MOUEZA, M.; GROS, O.; FRENKIEL, L. Embryonic, larval and postlarval development of the tropical clam, *Anomalocardia brasiliiana* (Bivalvia, Veneridae). **Journal of Molluscs Studies**. 65. 73-88. 1990.

NARCHI, W. Ciclo anual da gametogênese de *Anomalocardia brasiliiana* (Gmelin, 1791) (Mollusca: Bivalvia). **Bolm. Zool.**, São Paulo, p. 331-350, 1976.

NICOLAS, L., ROBERT, R. The effect of supply on metamorphosis and post-larval development in hatchery-reared *Pecten maximus*. **Aquaculture**. 192, 347-359. 2001.

OLIVEIRA, I.; AMORIM, A.; LAVANDER, H.; PEIXOTO, S. & GÁLVEZ, A.O. Spatial and temporal distribution of the shellfish *Anomalocardia brasiliiana* (Gmelin, 1791) on Mangue Seco beach, Pernambuco, Brazil. **Int. J. Aquat. Sci.**, v.2, n.1, p.68-79, 2011.

OLIVEIRA, I.B.; SILVA NETO, S.R.; LIMA FILHO, J.V.M.; PEIXOTO, S.R.M.; GÁLVEZ, A.O. Efeito do período chuvoso na extração do molusco bivalve *Anomalocardia brasiliiana* (Gmelin, 1791). **Agrária - Revista Brasileira de Ciências Agrárias**. v.9, n.1, p.139-145. 2014.

OLIVEIRA, R.L.M.; SOUZA A.B.; LAVANDER H.D.; ANTONIO I.G.; GUIMARÃES I.M.; CARDOSO JUNIOR L.O; PEIXOTO S.R. M.; GALVEZ A.O. Acompanhamento das fases de Crescimento da *Crassostrea rhizophorae* (Guilding, 1828), da larva ate seu tamanho comercial e testes de diferentes métodos de assentamento em laboratório. **Caminhos da Ciência**. Recife: ED. UFRPE. Vol. 2, p. 94-115. 2007.

PETTERSEN, A.K., TURCHINI, G.M., JAHANGARD, S., INGRAM, B.A., SHERMAN, C.D.H. Effects of different dietary microalgae of survival, growth, settlement and fatty acid composition of blue mussel (*Mytilus galloprovincialis*) larvae. **Aquaculture** (309): 115-124. 2010.

PEZZUTO, P.R., ECHTERNACHT, A.M. Avaliação de impactos da construção da via expressa SC-Sul sobre o berbigão *Anomalocardia brasiliiana* (Gmelin, 1791) (Mollusca: Pelecypoda) na reserve extrativista marinha de Pirajubaé (Florianópolis, SC-Brasil). **Atlântica** (21): 105-119. 1999.

PONIS, E., ROBERT, R., PARISI, G. Nutritional value of fresh and concentrated algal diets for larval and juvenile Pacific oyster (*Crassostrea gigas*). **Aquaculture**. 221, 491–505. 2003a.

PONIS, E., ROBERT, R., PARISI, G., TREDICI, M. Assessment of the performance of Pacific oyster (*Crassostrea gigas*) larvae fed with fresh and preserved *Pavlova lutheri* concentrates. **Aquaculture International**. 11, 69–79. 2003b.

PONIS, E., PROBERT, I., VE'RON, B., MATHIEU, M., ROBERT, R. New microalgae for the Pacific oyster *Crassostrea gigas* larvae. **Aquaculture**. (253): 618–627. 2006.

PRADO, S.; ROMALDE, J. L.; BARJA, J. L. Review of probiotics for use in bivalve hatcheries. **Veterinary Microbiology**. 145. 187-197. 2010.

PRIEUR, D.; MÉVEL, G.; NICOLAS, J.L.; PLUSQUELLEC, A.; VIGNEULLE, M. Interactions between bivalve molluscs and bacteria in the marine environment. **Oceanogr. Mar. Biol. Annu. Rev.** 28, 277–352. 1990.

REITAN, K.I., BOLLA, S., OLSEN, Y. A study of the mechanism of algal uptake in yolk-sac larvae of Atlantic halibut (*Hippoglossus hippoglossus*). **Journal Fish Biology**. 44. 303-310. 1994.

RIOS, E. **Seashells of Brazil**. 2.Ed. Rio Grande. Fundação da Universidade do Rio Grande. 492 p. 1994.

RICO-VILLA, B., LE COZ, J.R., MINGANT, C., ROBERT, R. Influence of phytoplankton diet mixtures on microalgae consumption, larval development and settlement of the Pacific oyster *Crassostrea gigas* (Thunberg). **Aquaculture** (256): 377–388. 2006.

RIVERO-RODRÍGUEZ, S.; BEAUMONT, A.R.; LORA-VILCHIS, M.C. The effect of microalgal diets on growth, biochemical composition, and fatty acid profile of *Crassostrea corteziensis* (Hertlein) juveniles. **Aquaculture**. 263, 199–210. 2007.

ROBERT, R., MINER, P., NICOLAS, J.L. Mortality control of scallop larvae in the hatchery. **Aquaculture International**. 4. 305-313. 1996.

RODRIGUES, A.M.L. Ecologia populacional do molusco bivalve *Anomalocardia brasiliiana* (Gmelin, 1791) (Bivalvia, Veneridae) em praias da região estuarina do Rio Apodi/Mossoró - RN. 93 f. **Dissertação** (Mestrado em Ciência Animal: Área de concentração: Aqüicultura e Ecologia Pesqueira) – Universidade Federal Rural do Semi-Árido. 2009.

RUIZ-AZCONA, P., RODRÍGUEZ-SIERRA, R., Martín, J.B. Culture of coquina clam, *Donax trunculus*, larvae. **Aquaculture**. 139. 151–155. 1996.

RUPP, G.S., PARSON, G.J. Effects of salinity and temperature on the survival and byssal attachment of the lion's paw scallop *Nodipecten nodosus* at its southern distribution limit. **J. Exp. Mar. Biol. Ecol.** 309, 173-198. 2004.

SILVA-CAVALCANTI, J.S. & COSTA, M.F. Fisheries in protected and non-protected areas: is it different? The case of *Anomalocardia brasiliiana* at tropical estuaries of Northeast Brazil. **Journal of Coastal Research**, SI 56 (Proceedings of the 10th International Coastal Symposium), Lisbon, Portugal, p. 1454 - 1458. 2009.

SOUTHGATE, P.C., BEER, A.C. Hatchery and early nursery culture of the blacklip pearl oyster (*Pinctada margaritifera* L.). **J. Shellfish Res.** 16. 561–567. 1997.

TANG, B., LIU, G. WANG, T. ZHANG & J. XIANG. Effects of various algal diets and starvation on larval growth and survival of *Meretrix meretrix*. **Aquaculture** (254): 526–533. 2006.

- TAYLOR, J.E., GUNN, B., WILLIAMS, P., MCNICOLL, I. An investigation of techniques for the hatchery and nursery rearing of the king scallop, *Pecten maximus* (L.) – Proceedings of the 9th international pectinid workshop, Nanaimo B.C., Canada, April 22-27, 1993. Volume 2, pp. 104-108, **Canadian Technical Report of Fisheries and Aquatic Sciences** 2, 1994.
- TORKILDSEN, L., SAMUELSEN, O.B., LUNESTED, B.T., BERGH, Ø. Minimum inhibitory concentrations of chloramphenicol, florfenicol, trimethoprim/sulfadiazine and flumequine in seawater of bacteria associated with scallops (*Pecten maximus*) Larvae. **Aquaculture**. 185. 1-2. 2000.
- VOLKMAN, J.K., DUNSTAN, G.A., JEFFREY, S.W., KEARNEY, P.S. Fatty acids from microalgae of the genus *Pavlova*. **Phytochemistry**. 30, 1855–1859. 1991.
- YAN, X., ZHANG, G., YANG, F. Effects of diet, stocking density, and environmental factors on growth, survival, and metamorphosis of Manila clam *Ruditapes philippinarum* larvae. **Aquaculture**. 253. 350-358. 2006.

5. - Artigo científico

5. 1. - Artigo científico I

Artigo científico encaminhado a Revista Latin American Journal of Aquatic Research.

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Growth and survival of *Anomalocardia brasiliiana* larvae (Bivalvia: Veneridae) fed microalgal diets

ABSTRACT. Laboratory production of bivalve molluscs depends of the use of microalgae. For this reason, the aim of this study was to evaluate the effect of using different microalgal diets on growth and survival of *Anomalocardia brasiliiana* larvae for 15 days of cultivation, between the trochophore and pediveliger stages. The diets were evaluated in two separate experiments. The first experiment tested the microalgae *Isochrysis galbana* (Ig), *Phaeodactylum tricornutum* (Phaeo), *Chaetoceros calcitrans* (Cca) and the combinations (Ig + Cca), (Ig + Phaeo) and (Cca + Phaeo). The second tested the microalgae *C. calcitrans* (Cca), *Pavlova lutheri* (Pl) and the combination (Cca + Pl). The microalgae *I. galbana* when given alone had lower survival and growth when compared to other diets tested. The use of diatoms *P. tricornutum* and *C. calcitrans* in this bivalve diet alone or combined other microalgae achieved better values for survival and growth. The diet (Cca + Pl) showed higher data concerning the growth and survival of *A. brasiliiana* larvae ($261.67 \pm 9.64 \mu\text{m}$ and $31.50 \pm 0.87 \%$). The use of microalgae *C. calcitrans* combined with *P. lutheri* is recommended to provide a good development in *A. brasiliiana* larvae. Alone, microalgae *P. lutheri* and *C. calcitrans* no showed significant difference, however, there is a synergistic effect when they are used in combination. The use of a specific diet of *P. lutheri* combined with *C. calcitrans* is recommended to provide satisfactory performance in *A. brasiliiana* larvae, cultured from the trochophore stage to pediveliger.

Keywords: *Anomalocardia brasiliiana* microalgae, larvae, growth, clam, diet, pediveliger.

Efecto de las dietas de microalgas en crecimiento larvario y la supervivencia de *Anomalocardia brasiliiana* (Bivalvia: Veneridae)

RESUMEN. La producción de moluscos bivalvos en laboratorio depende del uso de microalgas. En este sentido, el objetivo de este estudio fue evaluar el efecto del uso de diferentes dietas de microalgas sobre crecimiento y supervivencia de larvas de *Anomalocardia brasiliiana* durante los 15 días de cultivo entre las fases trocófora y pediveliger. Las dietas fueron evaluadas en dos experimentos separados. En el primer experimento, se evaluaron las microalgas *Isochrysis galbana* (Ig), *Phaeodactylum tricornutum* (Phaeo), *Chaetoceros calcitrans* (Cca) y las combinaciones (Ig + Cca), (Ig + Phaeo) y (Cca + Phaeo). En el segundo experimento, las microalgas, *C. calcitrans* (Cca), *Pavlova lutheri* (Pl) y su combinación (Cca + Pl). En el primer experimento, las dietas que contienen *C. calcitrans* mostraron la mayor longitud de la concha al final del cultivo, con una diferencia significativa entre Cca e Ig, donde la longitud promedio final fue de $251,33 \pm 5,66 \mu\text{m}$ y $230,00 \pm 7,42 \mu\text{m}$, respectivamente. Para el segundo experimento, con la dieta (Cca + Pl) se obtuvieron los mejores resultados para crecimiento y supervivencia del cultivo de larvas de *A. brasiliiana* ($261,67 \pm 9,64 \mu\text{m}$ y $31,50 \pm 0,87 \%$). Cuando las microalgas se usan de manera aislada *P. lutheri* y *C. calcitrans* no presentan significativas diferencias. Sin embargo hay un efecto sinérgico cuando se utilizan en combinación. El uso de la dieta específica de la especie de *P. lutheri* combinada con *C. calcitrans*, se recomienda para proporcionar un rendimiento satisfactorio en las larvas de *A. brasiliiana*, cultivadas a partir de la etapa trocófora paseo pediveliger.

Palabras clave: *Anomalocardia brasiliiana*, microalgas, larvas, crecimiento, molusco, dieta, pediveliger

INTRODUCTION

In 2010, global production of marine molluscs reached 13.9 million tons, and 75.5% of all aquatic organisms produced by mariculture in the world were molluscs, mostly bivalves, particularly oysters, mussels, clams and cockles (FAO, 2012). In Brazil, mollusc culture is still limited to a few species including Mytilidae *Perna perna*, Ostreidae *Crassostrea gigas*, *Crassostrea rhizophorae* and *Crassostrea brasiliiana* and in the Pectinidae family *Nodipecten nodosus*. Considering the potential of various species for cultivation, opportunity exists for the diversification of aquaculture.

Brazilian northeast region has favorable climatic conditions for the development of mollusc culture, but hatcheries have been established for few native species and are limited mainly to oysters, *C. rhizophorae* and *C. brasiliiana*. There is a need to develop new technologies for aquaculture of other native species, such as the bivalve *Anomalocardia brasiliiana* (Gmelin, 1791).

Anomalocardia brasiliiana is a resource of great importance for Brazilian coastal communities (Silva-Cavalcanti & Costa, 2009; Oliveira *et al.*, 2011). Laboratory production of its seed could be a management tool for restocking the heavily exploited populations along the Brazilian coastline, as well as for diversification of mariculture.

One of the obstacles to establish successful larval cultures is the availability of an appropriate diet (Ponis *et al.*, 2006; Liu *et al.*, 2009; Pettersen *et al.*, 2010). Microalgae have been the main food source for larvae and seeds in bivalve hatchery (Helm & Bourne, 2004). In addition, microalgae can provide improved water quality by absorbing toxic nitrogen products (ammonia and nitrite) and combating pathogenic bacteria for produce antibiotic substances (Lavens & Sorgeloos, 1996).

It is important that the microalgae are of the appropriate size to allow ingestion, and that they meet the nutritional requirements of the cultivated species. The cost of the microalgae must also be competitive with other species and be easy to produce on a commercial scale (Heasman *et al.*, 2000; Ponis *et al.*, 2008). Most microalgae used in bivalve hatcheries are species from the genera *Isochrysis*, *Pavlova*, *Chaetoceros*, *Phaeodactylum*, *Skeletonema*, *Thalassiosira*, *Dunaliella*, *Nannochloris*, *Tetraselmis* and *Rhodomonas* (Coutteau & Sorgeloos, 1992; Moueza *et al.*, 1999; Brown, 2002; Helm & Bourne, 2004; Cragg, 2006; Ponis *et al.*, 2008; Liu *et al.*, 2009).

Precise knowledge of microalgae as well as proper management is essential to provide greater shellfish larvae survival and growth. There isn't studies on the use of

microalgae in hatcheries of this species, and the purpose of this study was to evaluate survival and growth of larvae of *A. brasiliiana* when fed different microalgal diets.

MATERIALS AND METHODS

A total of 500 adults longer than 20 mm were collected on the beach of Mangue Seco (07°49' 44,19"S, 035°50' 03,06"W), Igarassu municipality, 30 km from Recife, Pernambuco state, Brazil, transported to the Laboratory of Sustainable Mariculture (LAMARSU). and acclimated during 24 h in 500 liter tanks at 25 °C, 30 g L⁻¹ salinity and 6 mg L⁻¹ mean dissolved oxygen. .

After this period, they were fed twice daily a mixture of *Isochrysis galbana* and *Chaetoceros calcitrans* at a cell ratio of 1:1, with a total ration of 20x10⁴ cell. mL⁻¹ .day⁻¹. In the first experiment, after 10 days, spawning occurred spontaneously, and the fertilized eggs were filtered 50 µm mesh opening and kept in a 500 L tank. Yet to the second experiment, spawns occurred on the same day that the breeding arrived at the laboratory, through the induction, release of gametes and gradual temperature increase (1 °C.h⁻¹)

After 24 hours D-veliger larvae (n = 30) had an average length of 69.94 ± 1.54 mm and were stocked with an initial density of five larvae.mL⁻¹, in triplicate for each treatment, in plastic containers with two liters of seawater (three µm filtered and UV-treated).

Temperature and salinity were measured daily, oxygen twice a day and the water of larval cultures was renewed completely every three days. Feeding was once a day, and microalgae supplied to the larvae were from three days-old cultures, in exponential growth.

The microalgae used in the experiment were obtained from LAMARSU stock strains. The seawater with salinity of 32 ± 2 g L⁻¹ was filtered through paper with 1 µm pores, autoclaved, and enriched with a Conway sterilized medium (Walne, 1966), supplemented with sodium metasilicate (40 mg L⁻¹) to provide a silica source for the diatom *C. calcitrans* and *P. tricornutum*.

The effect of microalgal diets on larval growth of *A. brasiliiana* was evaluated in two completely randomized experiments. The first experiment tested the microalgae *I. galbana* (Ig), *Phaeodactylum tricornutum* (Phaeo) and *Chaetoceros calcitrans* (Cca) and the combinations (Ig + Cca), (Ig + Phaeo) and (Cca + Phaeo). The larval rearing

period was 15 days, starting with the D-veliger larval stage and ending with pediveliger larvae. The total algal cell density provided was 30×10^3 cell mL⁻¹ and for bialgal diets a 1:1 ratio was used.

The second experiment evaluated the algae *C. calcitrans* (Cca), *P. lutheri* (Pl) and the combination (Cca + Pl). *C. calcitrans* was assessed again because it had the highest relative growth in the first experiment and was characterized as an excellent diet for weight gain in molluscs (Rivero-Rodriguez *et al.*, 2007). The microalgae *P. lutheri* has been described in some studies (Ponis *et al.*, 2006, 2008) as an excellent diet for bivalve molluscs. The methodology and algal density provided in the second experiment were the same adopted for the first experiment.

To assess larval survival at the end of experiment the entire volume of each experimental unit was filtered. The larvae were concentrated in a 50 mL container, from which one milliliter (1 mL) samples were drawn. The larval counting was done with a Sedgewick-Rafter counting chamber and optical microscope, three samples of each experimental unit were analyzed.

For the evaluation of larval growth, one milliliter samples of each experimental units were removed on the first and last day of the experiment and images of 30 larvae chosen at random were obtained using an optical microscope coupled to a camera lens, and their length (maximum anterior-posterior dimension) and width (maximum dorsal-ventral dimension) were measured using version 2.0 software ImageTool (Texas University, Health Science Center, San Antonio, USA)..

Relative growth (K) was calculated using the equation $K = (\ln L2 - \ln L1) / t$ (Walen, 1963), where *L1*, *L2* represent the lengths at the beginning and end of the experiment in µm, respectively, and *t* is the duration of the experiment in days.

Survival data, length, width and relative growth between diets, generated in both experiments, were previously checked for normality using the Kolmogorov-Smirnov test and for homogeneity of variance with Cochran's C test. Analysis of Variance (ANOVA) was used to determine the effect of diets on the growth and survival of larvae over time of cultivation. Duncan's test was performed to detect the mean levels which differed significantly between treatments. The level of significance was $P < 0.05$.

RESULTS

The variables of temperature (°C), salinity (g L⁻¹) and dissolved oxygen (mg L⁻¹) of the water were maintained within acceptable limits for shellfish growing. The temperature ranged from 25.05 °C to 25.60 °C, salinity from 29.88 g L⁻¹ to 30.18 g L⁻¹ and dissolved oxygen had a minimum value of 5.89 mg L⁻¹ and a maximum of 6.66 mg L⁻¹.

First Experiment

There was a significant difference in larval survival of *A. brasiliiana* as assessed at the end of cultivation, between diets (ANOVA). The Phaeo diet had the highest survival of approximately 25%, which was not significantly different of Cca, Ig + Phaeo and Ig + Cca diets. The lowest survival values were found in diets of Ig and Cca + Phaeo which averaged 6.83% and 5.27%, respectively, significantly lower than the Phaeo diet (Table 1).

No significant differences in shell width was found in larvae fed with the different algal diets tested (Table 1). Significant differences were observed between the relative growth rate in the diets tested (Table 1). The Cca and Ig + Phaeo diets had higher relative growth, than Ig diet.

Larval survival obtained in the first experiment reached a maximum value of 24.83 ± 3.03 % in the Phaeo diet. The survival of larvae fed with *C. calcitrans* (12.33%) was not significantly different from other diets tested (5.2- 4.83%). Larvae fed with microalgae *I. galbana* and a mixture of *C. calcitrans* with *P. tricornutum* had the lowest survival.

By analyzing the larval length as a growth factor, it was possible to determine significant differences between the diets tested ($P < 0.05$). The Cca diet had the greatest shell length at the end of cultivation (251.33 ± 5.66 µm), differing significantly from the Ig diet, which had the lowest growth in length (230.00 ± 7.42 µm). The other tested diets showed no significant difference regarding the growth in length (Fig. 1).

The microalgae *I. galbana* when given alone had lower survival and growth when compared to other diets tested. The use of diatoms *P. tricornutum* and *C. calcitrans* in this bivalve diet alone or combined other microalgae achieved better values for survival and growth.

Insert Table 1.

Insert Figure 1

Second experiment

Larval survival of *A. brasiliiana* assessed at the end of cultivation was significantly different among the three diets tested (Table 2). The Cca + Pl diet had the highest survival with a mean of 31.50%.

When analyzing the width of the shell as a growth parameter, no significant differences were found between the larvae fed with Cca + Pl and Cca diets (Table 2). The combined use of microalgae *C. calcitrans* and *P. lutheri* led to the highest growth in width after 15 days of culture with $242.00 \pm 10.02 \mu\text{m}$.

There were significant differences in the relative growth rate in larva with different diets (Table 2). The Cca + Pl diet showed higher relative growth significantly different than the Cca diet ($P < 0.05$), demonstrating its positive effect on the growth of *A. brasiliiana* larvae when microalgae *C. calcitrans* and *P. lutheri* are used in combination. Confirming the positive effect observed in larval survival, with a Cca + Pl diet.

By analyzing the length of the larvae, significant differences were found between the diets tested ($P < 0.05$). The diet containing both algae *C. calcitrans* and *P. lutheri* had the highest growth in length of larvae at the end of the experiment, with a significant difference from the Cca diet (Fig. 2).

Insert Table 2.

Insert Figure 2.

DISCUSSION

The water parameters, oxygen, temperature and salinity remained within the limits recommended for bivalve culture during all the experimental period. The absorption efficiency of some microalgae species, due to the maintenance of water quality standards can explain good growth in bivalves (Grizzle *et al.*, 2001; Resgalla Jr. *et al.*, 2007).

The use of a closed recirculation system, with daily water exchange can lead to better survival of *A. brasiliiana* larvae. Liu *et al.* (2009) were able to increase survival to 68.0% by using a continuous water renewal system (50% daily and 100% every three days) when *Clinocardium nutallii* larvae were fed some diet for nine days of cultivation. Thus, the low survival rate (25%) observed in *A. brasiliiana* larvae may be related to the use of closed cultivation without daily renewal, but with 100% renewal every three days.

In addition to maintaining water quality, a second factor of great importance for successful development of shellfish larvae is the nutritional value of the diet. Lourenço (2006) revised the chemical composition of some marine microalgae species, *C. calcitrans* (34% protein, 16% fat and 6% carbohydrate), *P. tricornutum* (30% protein, 14% fat and 8.4% carbohydrate), *I. galbana* (29% protein, 23% fat and 12.9% carbohydrate) and *P. lutheri* (29% protein, 12% lipids and 9% carbohydrate), indicating similar chemical compositions. However, slight differences in the concentration of fatty acids, vitamins and amino acids supplied in the diet can compromise survival and larval development.

The relative growth of larvae fed with Cca and Ig + Phaeo diets were higher only than that of larvae fed an Ig diet, with no significant differences with the other diets tested. The microalga *C. calcitrans* is considered the most suitable for feeding bivalve larvae (Brown & Robet, 2002), not only because of its biochemical composition, but also because of its cell size, digestibility and absence of toxins (Pettersen *et al.*, 2010).

Studies have found that the use of the microalgae *P. tricornutum* for feeding other bivalves, causes slow growth (Epifanio *et al.*, 1981; Albentosa *et al.*, 1996; Rivero-Rodriguez *et al.*, 2007), because it is difficult to digest (Rivero-Rodriguez *et al.*, 2007), probably due to its lack of tryptophan (Epifanio *et al.*, 1981). Tang *et al.* (2006) achieved a relatively low growth in *Meretrix meretrix* larvae when fed *Phaeodactylum tricornutum* (0.0388) and *Pavlova viridis* (0.0361).

A diet composed of the microalgae *C. calcitrans* showed the longest shell at the end of the cultivation (251.33 ± 5.66 μ m). Rivero-Rodriguez *et al.* (2007), evaluating the growth of *Crassostrea corteziensis* seeds found significant growth when using *C. calcitrans* alone or in combination with other diets, which was up to twice the growth when this microalga is present in the diet. This growth was affirmed to be related to the fact that *C. calcitrans* contains high levels of arachidonic acid (ARA). These authors

confirmed that the mixed use of *C. calcitrans* and *C. muelleri* is the second best diet in terms of weight gain for *C. corteziensis*. Similar results were found by Liu *et al.* (2009)

In the second experiment in this study, there was an increase in survival of *A. brasiliiana* larvae when *C. calcitrans* and *P. lutheri* microalgae were used in combination, reaching average survival above 31%. Ponis *et al.* (2008), evaluating the effect of *P. lutheri* in *Crassostrea gigas* larvae obtained survival above 78% when *C. calcitrans* was added to the diet. This corroborates our findings that after 15 days of culture, larvae fed a bialgal Cc + Pl diet had better survival that significantly differed from those fed only monoalgal diets (Cca and Pl). Other studies have also achieved good results when adding other species of diatom microalgae (Epifanio, 1979; Romberger & Epifanio, 1981; Laing & Millican, 1986; O'Connor & Heasman, 1997), which has been attributed to better essential nutrient balance (Webb & Chu, 1983).

The relative growth for the Cca + Pl diet (0.0867) was higher than the other diets tested in both experiments. There was no significant difference in growth when the microalgae *P. lutheri* and *C. calcitrans*, were used singly, however there is a synergistic effect when they are used together. The final average length of *A. brasiliiana* larvae fed a combination of *P. lutheri* and *C. calcitrans* was significantly higher than larvae fed only *C. calcitrans*.

Bialgal diets are often used in feeding bivalve larvae, and it is common to combine species, using a flagellate with a diatom, to maximize growth and larval development (Galley *et al.*, 2010; Liu *et al.*, 2009; Martínéz-Fernandez & Southgate, 2007). The flagellated species commonly used are *Isochrysis galbana* and *Pavlova lutheri* and the diatoms include *Chaetoceros calcitrans*. Spolaore *et al.* (2006) affirm that the combination of different algae species offers a better nutritional balance and improves animal growth compared to a monoalgal diet, which agrees with the result of this experiment, which found that the bialgal diet (Cca + Pl) led to better growth at the end of cultivation.

In hatchery of the mollusc *Pinctada margaritifera*, Martínéz-Fernandez & Southgate (2007) had a higher growth rate in the D-larvae phase when cultured with the ternary combination of *Pavlova* sp./*Pavlova salina*/*I. galbana*. However, the growth rate was not significantly higher than those of larvae fed with the binary combination of *P. salina*/*I. galbana* which has been used as a "standard diet" for *P. margaritifera* larvae (Southgate & Beer, 1997). Martínéz-Fernandez & Southgate (2007) suggest that feeding a single species of microalgae to *P. margaritifera* larvae may be more practical during

the first 10 days in a hatchery. However, the addition of diatom microalgae to the diet increased growth rate and survival in umbonate larvae of *P. margaritifera* when compared to treatments without diatoms.

Protein and vitamin content are important factors for determining the nutritional value of microalgae. Furthermore, high amounts of polyunsaturated fatty acids (eg. eicosapentaenoic [EPA], arachidonic acid [AA] and docosahexaenoic acid [DHA]), (Hemaiswarya *et al.*, 2011), can lead to better growth and survival of larvae fed with the microalgae *P. lutheri*, which is rich in DHA/EPA, and *C. calcitrans*, which is used to increase vitamin levels (Hemaiswarya *et al.*, 2011).

Pavlova lutheri is used frequently in the diet of the reproductive stock of bivalves, such as oysters, clams, mussels and scallops (Hemaiswarya *et al.*, 2011). *C. calcitrans* is rich in fatty acids such as arachidonic acid (ARA) and vitamins, and is widely used in bivalve hatchery (Hemaiswarya *et al.*, 2011; Rivero-Rodriguez *et al.*, 2007).

This study found that monoalgal and bialgal diets present satisfactory results in terms of survival. Prymnesiophyceae *P. lutheri* is commonly used in aquaculture as live food for marine invertebrates (molluscs, crustaceans, zooplankton) and particularly for bivalves (larvae, juveniles and breeding stock) (Webb & Chu, 1983; Borowitzka, 1997; Wikfors & Onho, 2001; Brow, 2002; Rico-Villa *et al.*, 2006), but its use alone may present low growth when compared to use with other diatoms (Rico-Villa *et al.*, 2006; Ponis *et al.*, 2008). The microalga *C. calcitrans* is used in some studies as a reference diet that offers good results for growth and survival (Ponis *et al.*, 2006; Ponis *et al.*, 2008).

Our results confirm the potential of the microalgae *C. calcitrans* for offering good growth of mollusc larvae. In hatching the mussel *Mytilus galloprovincialis*, was observed an increase in larval mortality when a diet of *C. calcitrans* was replaced with *C. muelleri*, with a consequent decrease in DHA content in the larvae (Pettersen *et al.*, 2010).

A comparison of the results of the first and second experiment in the current study indicated that the Cca + Pl diet had the best results in both survival ($31.50 \pm 0.87\%$) and growth ($261.67 \pm 9.64 \mu\text{m}$). The combination of the microalgae *C. calcitrans* and *P. lutheri* proved to be an excellent diet for the larval culture of *A. brasiliiana*. Used alone, the microalgae *C. calcitrans* and *P. lutheri* showed no significant difference, however, there is a synergistic effect when they are used in combination.

The use of microalgae *C. calcitrans* combined with *P. lutheri* can provide good growth in *Anomalocardia brasiliiana* larvae. The study found that the use of bialgal diets leads to better growth and survival, and is more recommended due to better nutritional balance. There is a need for new studies to improve dietary practices and increase survival between the larval stages, trochophore and pediveliger.

REFERENCES

- Albentosa, M., A. Pérez-Camacho, V. Labarta & M.J. Fernández-Reiriz. 1996. Evaluation of live microalgal diets for the seed culture of *Ruditapes decussatus* using physiological and biochemical parameters. *Aquaculture*. (148): 11-23.
- Boehs, G., T.M. Absher & A.C. da Cuz-Kaled. 2008. Ecologia populacional de *Anomalocardia brasiliiana* (Gmelin, 1791) (Bivalvia, Veneridae) na Baía de Paranaguá, Paraná, Brasil. *B. Inst. Pesca, São Paulo*, 34(2): 259 – 270.
- Borowitzka, M.A. 1997. Microalgae for aquaculture: Opportunities and constraints. *J. Appl. Phycol.* (9): 393-401.
- Brown, M.R., 2002. Nutritional value of microalgae for aquaculture. In: Cruz-Suárez, L. E., Ricque-Marie, D., Tapia-Salazar, M., Gaxiola-Cortés, M. G., Simoes, N. (eds.). *Avances en Nutrición Acuicola VI. Memorias del VI Simposium Internacional de Nutrición Acuicola. 3 al 6 de Septiembre del 2002. Cancún, Quintana Roo, México.*
- Brown, M. & R. Robert. 2002. Preparation and assessment of microalgal concentrates as feed for larval and juvenile Pacific oyster (*Crassostrea gigas*). *Aquaculture*. (207): 289-309.
- Coutteau, P. & P. Sorgeloos. 1992. The requirement for live algae and their replacement by artificial diets in the hatchery and nursery rearing of bivalve molluscs: an international survey. *J. Shellfish Res.* (11): 467-476.
- Cragg, S.M. 2006. Development, physiology, behaviour and ecology of scallop larvae. In: Shumway, S.E. & G.J. Parsons. (eds.), *Scallops: Biology, Ecology and Aquaculture*. Elsevier, B.V. Amsterdam. Ch. 2: 45-122 pp.
- Dias, T.L.P., R.S. Rosa & L.C.P. Damasceno. 2007. Aspectos socioeconômicos, percepção ambiental e perspectivas das mulheres marisqueiras da Reserva de Desenvolvimento Sustentável Ponta do Tubarão (Rio Grande do Norte, Brasil). *Gaia Scientia*. 1(1): 25-35.

- 369 Epifanio, C.E., 1979. Growth in bivalve molluscs: nutritional effects of two or more
370 species of algae in diets fed to the American oyster *Crassostrea virginica*
371 (Gmelin) and the hard clam *Mercenaria mercenaria* (L.). Aquaculture. (18): 1–12.
- 372 Epifanio, C.E., C.C. Valenti & C.L. Turk, 1981. A comparison of *Phaeodactylum*
373 *tricornutum* and *Thalassiosira pseudonana* as foods for the oyster *Crassostrea*
374 *virginica*. Aquaculture. (23): 247–253.
- 375 Espinosa, E.P. & B. Allam. 2006. Comparative growth and survival of juvenile hard
376 clams, *Mercenaria mercenaria*, fed commercially available diets. Zoo Biol.,
377 25:513–525.
- 378 FAO - Food and Agriculture Organization of the United Nations. 2012. The state of
379 world fisheries and aquaculture. Rome. 251 p.
- 380 Galley, T.H., F.M. Batista, R. Braithwaite, J. King & A.R. Beaumont. 2010.
381 Optimisation of larval culture of the mussel *Mytilus edulis* (L.). Aquacult. Int.
382 (18):315–325
- 383 Grizzle, R.E., V. M. Bricej & S.E. Shumway. 2001. Physiological ecology of
384 *Mercenaria mercenaria*. In: Kraeuter, J.N. & M. Castagna (eds), Biology of the
385 Hard Clam. Elsevier, B.V. Ch. 8: 305-333.
- 386 Helm, M.M. & N. Bourne. 2004. Hatchery culture of bivalve: a practical manual. FAO.
387 Fish. Tech. Pp. 471, Rome, 177 p.
- 388 Hemaiswarya, S., R. Raja, R. Ravi Kumar, V. Ganesan & C. Anbazhagan. 2011.
389 Microalgae: a sustainable feed source for aquaculture. World J. Microbiol.
390 Biotechnol. (27): 1737–1746.
- 391 Heasman M.P., J. Diemar, W. O'Connor, T. Sushames & L. Foulke. 2000.
392 Development of extended shelf-life microalgae concentrate diets harvested by
393 centrifugation for bivalve mollusks: a summary. Aquacult. Res. (31):6, 37–59.
- 394 Lavander, H.D., L.O. Cardoso Jr., R.L. Oliveira, S.R. Silva Neto, A.O. Gálvez &
395 S.R.M. Peixoto. 2011. Biologia reprodutiva da *Anomalocardia brasiliiana*
396 (Gmelin, 1791) no litoral norte de Pernambuco, Brasil. Rev. Bras. Ciênc. Agrárias
397 6(2): 344-350.
- 398 Lavens P. & P. Sorgeloos. 1996. Manual on the production and use of live food for
399 aquaculture. FAO Fisheries Technical paper. In: Lavens P, Sorgeloos P (eds)
400 Rome. 36–19 pp.
- 401 Laing I. & P.F. Millican. 1986. Relative growth and growth efficiency of *Ostrea edulis*
402 L. spat fed various algal diets. Aquaculture (54): 245–62.

- 403 Liu, W., C.M. Pearce, A.O. Alabi & H. Gurney-Smith. 2009. Effects of microalgal diets
404 on the growth and survive of larvae and pos-larvae of the basket cockle,
405 *Clinocardium nuttallii*. Aquaculture (293): 248-254.
- 406 Lourenço, S.O. 2006. Cultivo de microalgas marinhas – princípios e aplicações. São
407 Carlos: 606 p.
- 408 Martins, V.S. & F.J.B Souto. 2006. Uma análise biométrica de bivalves coletados por
409 marisqueiras no manguezal de Acupe, Santo Amaro, Bahia: uma abordagem
410 etnoconservacionista. Sitientibus Sér. Ci. Biol. (6): 98-105.
- 411 Martínez-Fernández, E. & P.C. Southgate. 2007. Use of tropical microalgae as food for
412 larvae of the black-lip pearl oyster *Pinctada margaritifera*. Aquaculture (263):
413 220–226.
- 414 Moueza, M., O. Gros & L. Frenkiel. 1999. Embryonic, larval and postlarval
415 development of the tropical clam, *Anomalocardia brasiliiana* (Bivalvia,
416 Veneridae). J Moll Stud. (65): 73-88.
- 417 Nishida, A.K., N. Nordi & R.R.N. Alves. 2004. Abordagem etnoecológica da coleta de
418 moluscos no litoral paraibano. Trop. Oceanogr. 32 (1): 53-68.
- 419 O'Connor, W.A. & M.P., Heasman. 1997. Diet and feeding regimens for larval
420 doughboy scallops *Mimachlamys asperima*. Aquaculture (158): 289–303.
- 421 Oliveira, I.; A. Amorim, H. Lavander, S. Peixoto & A. O. Galvéz. 2011. Spatial and
422 temporal distribution of the shellfish *Anomalocardia brasiliiana* (Gmelin, 1791)
423 on Mangue Seco beach, Pernambuco, Brazil. Int. J. Aquat. Sci. 2 (1), 68-79.
- 424 Pettersen, A.K., G.M. Turchini, S. Jahangard, B.A. Ingram & C.D.H. Sherman. 2010.
425 Effects of different dietary microalgae of survival, growth, settlement and fatty
426 acid composition of blue mussel (*Mytilus galloprovincialis*) larvae. Aquaculture
427 (309): 115-124.
- 428 Pezzuto, P.R. & A. M. Echternacht. 1999. Avaliação de impactos da construção da via
429 expressa SC-Sul sobre o berbigão *Anomalocardia brasiliiana* (Gmelin, 1791)
430 (Mollusca:Pelecypoda) na reserve extrativista marinha de Pirajubaé
431 (Florianópolis, SC-Brasil). Atlântica (21): 105-119.
- 432 Ponis, E., I. Probert, B. Ve´ron, M. Mathieu & R. Robert. 2006. New microalgae for the
433 Pacific oyster *Crassostrea gigas* larvae. Aquaculture. (253): 618–627.
- 434 Ponis, E., G. Parisi, G. Chini Zittelli, F. Lavista, R. Robert & M.R. Tredici. 2008.
435 *Pavlova lutheri*: Production, preservation and use as food for *Crassostrea gigas*
436 larvae. Aquaculture (282): 97–103.

- 437 Resgalla Jr., C., E.S. Brasil, K. Laitano & R.W. Reis Filho. 2007. Physioecology of the
438 mussel *Perna perna* (Mytilidae) in southern Brazil. *Aquaculture* (270): 464-474.
- 439 Rico-Villa, B., J.R. Le Coz, C. Mingant & R. Robert. 2006. Influence of phytoplankton
440 diet mixtures on microalgae consumption, larval development and settlement of
441 the Pacific oyster *Crassostrea gigas* (Thunberg). *Aquaculture* (256): 377–388.
- 442 Rivero-Rodriguez, S., A.R. Beaumont & M.C. Lora-Vilchis. The effect of microalgal
443 diets on growth, biochemical composition, and fatty acid profile of *Crassostrea*
444 *corteziensis* (Hertlein) juveniles. *Aquaculture* (263): 199-210.
- 445 Robert, R., G. Parisi, L. Rodolfi, B.M. Poli & M.R. Tredici. 2001. Use of fresh and
446 preserved *Tetraselmis suecica* for feeding *Crassostrea gigas* larvae. *Aquaculture*
447 (192): 333–346.
- 448 Romberger, E.P. & C.E Epifanio. 1981. Comparative effects of diets consisting of one
449 or two algal species upon assimilation efficiencies and growth of juvenile oysters,
450 *Crassostrea virginica* (Gmelin). *Aquaculture* (25): 77–87.
- 451 Silva-Cavalcanti, J.S. & M.F. Costa. 2009. Fisheries in protected and non-protected
452 areas: is it different? The case of *Anomalocardia brasiliiana* at tropical estuaries of
453 northeast Brazil. *J Coastal Res. SI 56* (Proc. 10th Int. Coastal Symp.): 1454 –
454 1458.
- 455 Southgate, P.C. & A.C. Beer. 1997. Hatchery and early nursery culture of the blacklip
456 pearl oyster (*Pinctada margaritifera* L.). *J Shellfish Res.* (16): 561–567.
- 457 Spolaore, P., C. Joannis-Cassan, E. Duran & A. Isambert. 2006. Commercial
458 applications of microalgae. *J Biosci Bioeng* (101): 87–96.
- 459 Tang, B., B. Liu, G. Wang, T. Zhang & J. Xiang. 2006. Effects of various algal diets
460 and starvation on larval growth and survival of *Meretrix meretrix*. *Aquaculture*
461 (254): 526–533.
- 462 Webb, K.L. & F.L. Chu. 1983. Phytoplankton as a source of food for bivalve larvae. In:
463 Pruder, G.D., Langdon, C., Conklin, D. (eds.), *Proceedings of the 2nd*
464 *International Conference on Aquaculture Nutrition: Biochemical and*
465 *physiological approaches to shellfish nutrition.* October 1981, Rehoboth Beach,
466 Delaware. Louisiana State University Press, Baton Rouge. 272–291 pp.
- 467 Wikfors G.H. & M. Ohno. 2001. Impact of algal research in aquaculture. *J Phycol.* (37):
468 968–974.
- 469 Wilson, J.H., 1978. The food value of *Phaeodactylum tricornutum* Bohlin to the larvae
470 of *Ostrea edulis* L. and *Crassostrea gigas* Thunberg. *Aquaculture* (13): 313– 323.

Table 1. Mean ($\pm$ SE) survival, width and relative growth (K) of larvae of *A. brasiliiana* fed different diets over 15 days of culture. Cca: *Chaetoceros calcitrans*, Ig: *Isochrysis galbana*; Phaeo: *Phaeodactylum tricornutum*, mixed diets Cca+Ig: *C. calcitrans* and *I. galbana*; Cca + Phaeo: *C. calcitrans* and *P. tricornutum*, and Ig + Phaeo: *I. galbana* and *P. tricornutum*. Different letters in the same column have a significant difference ($P < 0.05$).

Diets	Survival (%)	Width (μm)	K ($\mu\text{m}.\text{day}^{-1}$)
Ig	6.83 ± 1.92^b	212.33 ± 6.79	0.0779 ± 0.0022^b
Cca	12.33 ± 5.84^{ab}	229.00 ± 5.19	0.0848 ± 0.0015^a
Phaeo	24.83 ± 3.03^a	222.33 ± 5.27	0.0830 ± 0.0020^{ab}
Ig + Cca	13.50 ± 4.91^{ab}	226.33 ± 5.04	0.0821 ± 0.0020^{ab}
Ig + Phaeo	18.27 ± 2.42^{ab}	226.00 ± 7.81	0.0839 ± 0.0019^a
Cca + Phaeo	5.27 ± 1.36^b	220.00 ± 6.49	0.0807 ± 0.0020^{ab}

Table 2. Mean ($\pm$ SE) survival, width and relative growth (K) of larvae of *A. brasiliiana* fed different diets over 15 days of culture. Cca: *Chaetoceros calcitrans*, Pl: *Pavlova lutheri*, mixed diet Cca+Pl: *C. calcitrans* and *P. lutheri*. Different letters in the same column have a significant difference ($P < 0.05$).

Diets	Survival (%)	Width (μm)	K ($\mu\text{m}.\text{day}^{-1}$)
Cca	4.42 ± 1.58^c	211.67 ± 8.60^b	0.0781 ± 0.0028^b
Pl	16.80 ± 5.63^b	219.33 ± 6.69^{ab}	0.0805 ± 0.0021^{ab}
Cca+Pl	31.50 ± 0.87^a	242.00 ± 10.02^a	0.0867 ± 0.0024^a

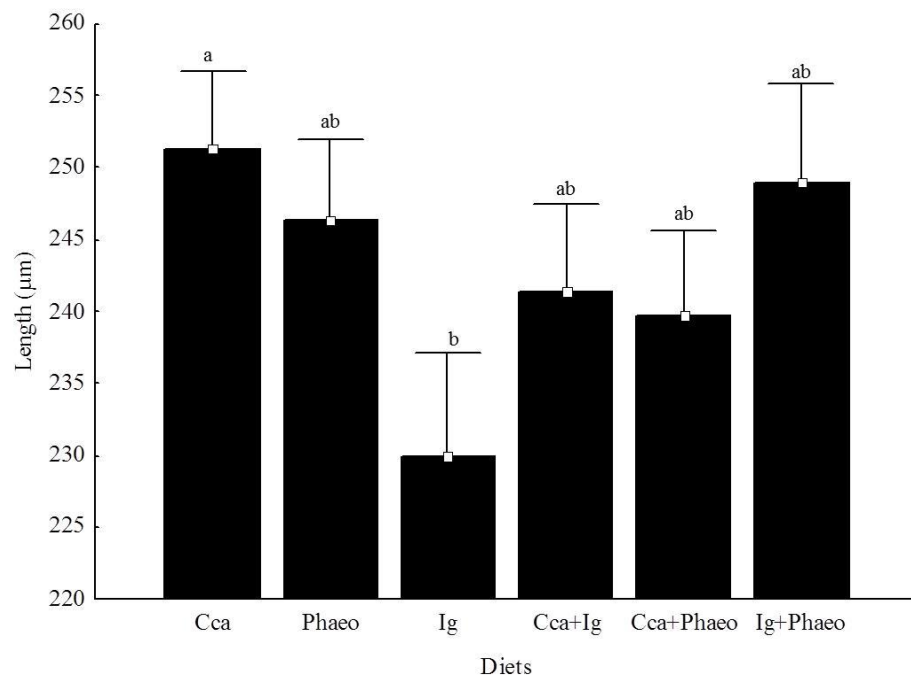


Figure 1. Mean length of larvae *A. brasiliana* fed different diets of single or mixed algae over a period of 15 days of growth. Cca: *Chaetoceros calcitrans*, Ig: *Isochrysis galbana*; Phaeo: *Phaeodactylum tricornutum*, mixed diets of Cca + Ig: *C. calcitrans* and *I. galbana*; Cca + Phaeo: *C. calcitrans* and *P. tricornutum*, and Ig+Phaeo: *I. galbana* and *P. tricornutum*. Means with different letters differ significantly ($P < 0.05$).

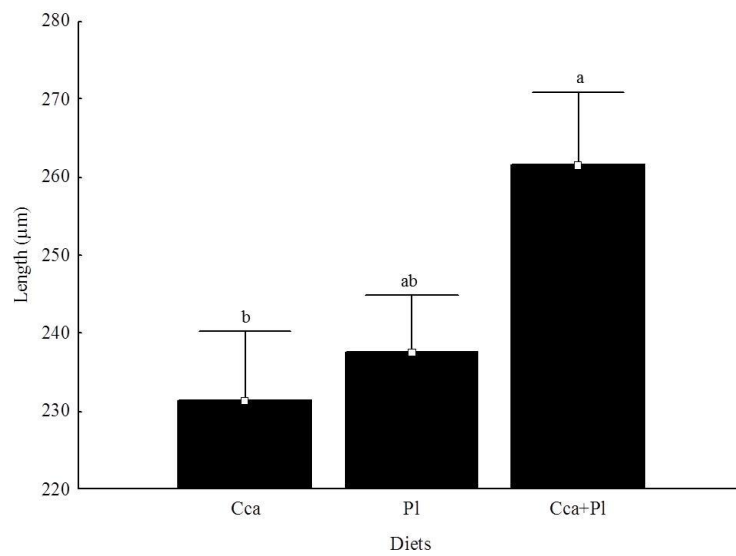


Figure 2. Mean length of larvae of *A. brasiliiana* fed microalgal diets for a period of 15 days. Ig: *Isochrysis galbana*; Cca: *Chaetoceros calcitrans*; Pl: *P. lutheri*. Means with different letters have a significant difference ($P < 0.05$).

5.2. - Artigo científico II

Artigo científico a ser encaminhado a Revista Aquaculture.

Todas as normas de redação e citação, deste capítulo, atendem as estabelecidas pela referida revista (em anexo).

Efeito da densidade de estocagem no crescimento e sobrevivência de pós-larvas de
Anomalocardia brasiliana (Bivalvia: Veneridae)

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Resumo

As pós-larvas de *Anomalocardia brasiliiana* foram cultivadas por 28 dias para avaliação do efeito da densidade de estocagem no crescimento e sobrevivência, sendo alimentados com a mistura das microalgas *Pavlova lutheri* e *Chaetoceros calcitrans*. Três densidades de estocagem foram testadas 40, 80 e 160 ind.cm⁻², em unidades experimentais de pequeno volume (2 litros) com sistema de cultivo estático e renovação total da água feita a cada 48h. O delineamento experimental utilizado foi inteiramente casualizado com três tratamentos (densidades) e três repetições cada. Os animais cultivados na densidade de estocagem de 40 ind.cm⁻² alcançaram 1mm de comprimento aos 24 dias de cultivo. Ao final dos 28 dias de cultivo apenas 18% dos animais cultivados na densidade de 80 ind.cm⁻² apresentavam comprimento de 1mm, enquanto que o tratamento com 160 ind.cm⁻² 100% não atingiram 1mm de comprimento. Ao avaliarmos todo o período experimental (0-28 dias) a densidade de 40 ind.cm⁻² apresentou a maior taxa de crescimento específico diária, 4,98±0,08 %.dia⁻¹. A taxa de sobrevivência das pós-larvas para as menores densidades (40 e 80 ind.cm⁻²) apresentaram maiores médias (53,24 ± 4,60 % e 52,95 ± 3,32 %) respectivamente, diferindo significativamente da maior densidade de estocagem com 31,54 ± 0,70 %. Na larvicultura das pós-larvas de *A. brasiliiana* deve-se realizar o manejo da densidade de estocagem no decorrer do crescimento deste molusco. A densidade de 160 ind.cm⁻² pode ser utilizada até que as pós-larvas alcancem 600µm de comprimento, pós-larvas maiores que 600 µm devem ser cultivadas na densidade de 40 larvas.cm⁻² para manter a taxa de crescimento máximo diária.

Palavras-chave: crescimento, sobrevivência, pós-larvas, larvicultura.

1. Introdução

O marisco, *Anomalocardia brasiliiana*, é um recurso pesqueiro de grande importância para os estados brasileiros, principalmente, nas comunidades costeiras da região nordeste (Nishida et al., 2004; Silva-Cavalcanti & Costa, 2009; Oliveira et al., 2011; Lavander et al., 2011; Oliveira et al., 2014). Pertence à família Veneridae, vive em profundidades de 0,5 a 1,5 m, superficialmente escavado na areia próximo ao manguezal, é sensível a variações ambientais, com alta mortalidade devido às chuvas, que causam grandes flutuações no tamanho e na distribuição das populações (Monti et al., 1991; Mouëza et al., 1999; Oliveira et al., 2014).

A região nordeste do Brasil apresenta condições climáticas favoráveis para o desenvolvimento da malacocultura, no entanto para as espécies nativas ainda é pouco desenvolvida a larvicultura. A espécie *A. brasiliiana* apresenta potencial para ser cultivada, contudo o estabelecimento de um protocolo de produção larval para uma nova espécie exige a avaliação de diversos fatores tais como: densidade de estocagem, qualidade e quantidade da dieta a ser fornecida, e parâmetros de qualidade da água de cultivo (temperatura e salinidade).

Foram realizados diversos estudos para estabelecimento de um protocolo de produção de larvas de moluscos, tais como: bivalves sedentários (Liu et al, 2002; Liu et al, 2006; Yan et al, 2006; Liu et al, 2008; Liu et al, 2010), mexilhões (Galley et al, 2010; Pettersen et al, 2010), ostras (Rico-Villa et al, 2006) e vieiras (Rupp e Parsons, 2004; Gouda et al, 2006). Observou-se que os fatores que influenciam o crescimento e a sobrevivência das larvas é essencial para o sucesso de uma larvicultura.

Uma importante variável a ser considerada é a densidade de estocagem, pois afeta no crescimento e sobrevivência de larvas, juvenis e adultos bivalves tem sido amplamente estudado (Fréchette, 2005), principalmente para aquelas espécies com

importância comercial para aquicultura. A maioria dos estudos avaliaram o período de crescimento nas fases de D véliger até a metamorfose para pé de véliger (Liu et al., 2006; Yan et al., 2006; Liu et al., 2010), pois são fases críticas, onde as larvas são frágeis e susceptíveis a doenças durante o manejo.

Quando as larvas realizam metamorfose para a etapa pé de véliger, ficam mais resistentes ao manejo e passam a ser cultivadas em densidades mais elevadas, por um período maior de tempo (semanas) até alcançarem o tamanho de semente (1mm) quando poderão ser transferidas para o campo (Nicolas e Robert, 2001; Epelbaum et al., 2011; Liu et al., 2011). A Avaliação da densidade de estocagem durante a fase pós-larval tem como objetivo promover um ambiente de cultivo adequado para que estas alcancem a fase de semente em um menor tempo possível.

Uma larvicultura comercial tem por meta melhorar os dados de cultivo, aumentando a produção em um volume limitado de água com o menor custo possível. Nas larviculturas de moluscos é comum utilizar os sistemas de fluxo contínuo de água, diferenciados pelo sentido da corrente, de cima para baixo (Down-welling) e de baixo para cima (Up-welling). Contudo, quando se utiliza unidades de pequeno volume, o sistema de cultivo fechado com circulação da água também é bastante utilizado.

Neste sentido, este estudo tem como objetivo avaliar o efeito da densidade de estocagem sobre o crescimento e a sobrevivência de pós-larvas de *Anomalocardia brasiliiana* em um sistema de cultivo fechado.

2. Material e Métodos

Para obtenção de larvas de *A. brasiliiana* foi necessário à coleta de indivíduos adultos no ambiente natural. Os reprodutores foram capturados na praia de Mangue Seco (07° 49' 44,19"S e 035° 50' 03,06"O), litoral norte do estado de Pernambuco –

Brasil. Foram coletados 500 indivíduos adultos, com comprimento médio de $24,50 \pm 2,00$ mm e transportados ao Laboratório de Maricultura Sustentável (LAMARSU). No laboratório os animais foram aclimatados em tanques de 500 litros com controle das variáveis de qualidade da água, temperatura ($25\text{ }^{\circ}\text{C}$), salinidade (30 g.L^{-1}) e oxigênio dissolvido médio de (6 mg.L^{-1}). Após a identificação dos mariscos que estavam com as gônadas maduras foram realizados estímulos para desova tais como: aumento da temperatura, oferta de alimento e lançamento de gametas na água. Após estes estímulos foram obtidas as desovas. Os ovos fecundados foram filtrados através de uma malha com abertura de $50\text{ }\mu\text{m}$ e mantidos em incubadoras com 30L de volume útil por 15 dias, com troca total da água a cada 48h. Durante esse tempo as larvas foram alimentadas com uma mistura das microalgas *Pavlova lutheri* e *Chaetoceros calcitrans*, na proporção celular de 1:1 e a concentração algal foi diferenciada para cada estágio larval e está descrito na Tabela 1.

Inserir Tabela 1

Larvas pé de véliger com 15 dias de vida foram mensuradas ($n=30$), obtendo-se um tamanho médio de $307,89 \pm 50,92\text{ }\mu\text{m}$ de comprimento (medida entre as extremidades ântero-posterior da concha). Em seguida, foram transferidas para as unidades experimentais que eram constituídas de recipientes plásticos transparentes com dois litros de volume útil, internamente suspenso um cilindro de PVC (diâmetro 10 cm; área $78,5\text{ cm}^2$), sendo na base do cilindro colocado uma malha de $250\text{ }\mu\text{m}$ ficando a uma distância de fundo de 5 cm e adicionado um airlift para circulação da água (Figura 1).

Inserir Figura 1

As pós-larvas foram cultivadas por um período de 28 dias, com avaliação de três diferentes densidades de estocagem, 40 ind.cm⁻², 80 ind.cm⁻² e 160 ind.cm⁻², em um sistema fechado de circulação da água com troca total a cada 48 horas. A água utilizada na larvicultura foi previamente esterelizada por radiação ultravioleta, para posterior utilização nas unidades experimentais. Diariamente foram aferidas as variáveis temperatura, salinidade e oxigênio dissolvido. A qualidade da água nas unidades experimentais foi mantida dentro dos limites aceitáveis para a larvicultura de bivalves, com temperatura média de 25,89±0,50 °C, salinidade média de 28,5±0,88 g.L⁻¹ e com média de oxigênio dissolvido em 5,66 ±0,55 mg.L⁻¹).

A alimentação foi realizada uma vez ao dia, com as microalgas *P. lutheri* e *C. calcitrans*, fornecidas às pós-larvas a partir do terceiro dia de cultivo algal, na fase exponencial da curva de crescimento, a uma concentração de 50x10³ cel.mL⁻¹. As microalgas foram obtidas do banco de cepas do LAMARSU, e mantidas em tubos de ensaio com água marinha de salinidade 32±2 g.L⁻¹ e enriquecida com meio Conway esterelizado, sendo adicionado metassilicato de sódio (40 mg.L⁻¹) como fonte de sílica no cultivo da diatomácea *C. calcitrans*.

Para a avaliação do crescimento larval, foram retiradas amostras de um mililitro das unidades experimentais e com auxílio da câmara de Sedgewick-Rafter, imagens foram tomadas utilizando uma câmara fotográfica objetiva acoplada a um microscópio óptico. Foram realizadas fotografias aleatórias em 30 pós-larvas de cada amostra, a cada semana. As medidas de comprimento (máxima dimensão antero-posterior) e largura (máxima dimensão dorso-ventral) das larvas foram obtidas utilizando o software ImageTool versão 2.0 (Texas University, Health Science Center, San Antonio, USA).

Durante o experimento a taxa de crescimento das pós-larvas foram estimadas usando a seguinte fórmula:

$$TCE = 100 (\ln C2 - \ln C1) / t,$$

Onde TCE é a taxa de crescimento específico (% dia⁻¹); C1, C2 representam os comprimentos no início e no fim do experimento em µm, respectivamente, e t é o tempo de duração do experimento (dias).

Para a avaliação da sobrevivência larval no final do cultivo, foi realizada uma filtragem de todo o volume de cada unidade experimental. As pós-larvas foram concentradas em recipientes de 50 mL, e retiradas amostras de um mililitro (mL). A contagem dos indivíduos foi realizada com auxílio da câmara Sedgewick-Rafter e microscópio óptico, e analisado três amostras de cada unidade experimental.

Os dados de sobrevivência, comprimento e crescimento das densidades de estocagem avaliadas, foram previamente checados quanto à normalidade dos dados usando o teste de Kolmogorov-Smirnov e para homogeneidade de variância com o teste de Cochran C. A Análise de Variância (ANOVA) foi usada para determinar o efeito das densidades no crescimento e sobrevivência das pós-larvas ao longo do tempo de cultivo. O teste de Tukey foi realizado para detectar qual média entre os tratamentos diferem significativamente, a nível de significância de 5% (p< 0,05). Os dados estão apresentados em média ± erro padrão.

3. Resultados

O comprimento médio das pós-larvas de *A. brasiliiana* estocadas em diferentes densidades de estocagem, estão apresentados na Figura 2.

Ao término da primeira semana do experimento (7 dias) não foram encontradas diferenças (p>0,05) no comprimento dos animais entre as densidades testadas. A partir

da segunda semana o comprimento das pós-larvas foram menores a medida que se aumentou a densidade de estocagem. Aos 21 dias de cultivo não foi identificado diferença ($p>0,05$) no comprimento das pós-larvas cultivadas com as densidades 40 ind.cm⁻² e 80 ind.cm⁻². As pós-larvas de *A. brasiliiana* cultivadas na maior densidade de estocagem (160 ind.cm⁻²) apresentaram menor média de comprimento ($p<0,05$), tornando-se mais evidente ao final dos 28 dias de cultivo.

As pós-larvas cultivadas na densidade de 40 ind.cm⁻² alcançaram nos 24 dias de cultivo 1mm de comprimento, e ao final dos 28 dias do experimento apenas 18% das larvas cultivadas na densidade de 80 ind.cm⁻² apresentavam comprimento de 1mm, enquanto que no tratamento com 160 ind.cm⁻² não atingiram este comprimento.

Inserir Figura 2

A tabela 2 apresenta a TCE (% dia⁻¹) das três diferentes densidades de estocagem para cada período do cultivo. O rápido crescimento ocorreu para os primeiros sete dias do experimento. A menor densidade (40 ind.cm⁻²) continuou apresentando o maior crescimento até 14 dias de cultivo, diferindo significativamente das outras densidades avaliadas ($p<0,05$). Aos 21 dias de cultivo as pós-larvas de todos os tratamentos apresentaram uma queda acentuada na taxa de crescimento (Tabela 2).

As pós-larvas cultivadas na densidade de 40 ind.cm⁻² apresentam a maior taxa de crescimento, diferindo significativamente das outras densidades testadas. Ao avaliarmos todo o período experimental (0-28 dias) a densidade de 40 ind.cm⁻² permaneceu com a maior taxa de crescimento específico diária.

Inserir Tabela 2

A sobrevivência larval das menores densidades (40 e 80 ind.cm⁻²) apresentaram média de 53,24 ± 4,60 % e 52,95 ± 3,32 % respectivamente, diferindo significativamente da maior densidade de estocagem com 31,54 ± 0,70 % (Figura 3). Portanto, observa-se o efeito negativo na sobrevivência quando ocorre aumento na densidade de estocagem de pós-larvas de *A. brasiliiana*.

Inserir Figura 3

4. Discussão

O alimento é um fator limitador do crescimento, de larvas de moluscos bivalves quando são cultivadas em altas densidades. Estudos evidenciam que (Loosanoff e Davis, 1963; Sprung, 1984a,b), o excesso de alimento pode causar um gasto energético com a produção de pseudofazes e outros efeitos inibitórios relacionados à fisiologia de digestão, resultando em uma diminuição do crescimento larval, além de ocasionar contaminação bacteriana e toxidez alimentar pela acumulação de ectometabolitos (Loosanoff e Davis, 1963; Sprung, 1984a,b).

Ao estudar o efeito da densidade de estocagem de uma espécie-alvo deve se garantir que o alimento não ultrapasse a concentração máxima desejável (Liu et al., 2010). No presente estudo, o residual algal foi verificado diariamente, sendo ajustado quando necessário e mantido na mesma concentração da dieta em cada tratamento. Desta forma, supõe-se que o alimento não tenha sido um fator limitante.

Altas densidades de estocagem no cultivo de larvas de bivalves podem ocasionar redução no crescimento, isto pode ser atribuído além da escassez do alimentação, também ao reduzido espaço de confinamento que causará deterioração na qualidade da

226 água (Reghavan e Gopinathan, 2008; Velasco e Barros, 2008). Larvas pelágicas
227 utilizam o velum para nadar e se alimentar, logo um aumento na densidade de cultivo
228 aumenta a possibilidade de colisão entre os indivíduos, que pode resultar numa inibição
229 da atividade alimentar (Liu et al., 2006).

230 Na metamorfose de larvas pelágicas para pé de véliger, quando estas passam a ter
231 hábitos bentônicos, há uma queda na taxa de crescimento (Liu et al., 2006). Além do
232 fator biológico, estudos evidenciam que a área de fundo pode ser um fator limitante de
233 densidade, para larvas com hábitos bentônicos. Em larvas pé de véliger há uma redução
234 no crescimento quando os indivíduos ocupam 100% ou mais da área de cobertura de
235 fundo (Liu et al., 2011). Heasman et al. (2002) observou que ocupar 70% da área de
236 cobertura de fundo na larvicultura de juvenis de vieiras *Pecten fumatus* é um fator
237 limitador da densidade para garantir a manutenção da máxima taxa de crescimento.

238 Usando a área de fundo (100%) como fator limitador da densidade para larvas pé
239 de véliger de *Clinocardium nuttallii*, foi possível determinar que 160 ind.cm⁻² seria o
240 recomendado para indivíduos com até 1mm de comprimento (Liu et al., 2011). O fator
241 área de cobertura de fundo não foi limitador de crescimento para *A. brasiliiana*, pois
242 utilizando a metodologia de Liu et al. (2011) obtivemos ao final do cultivo 26,74 %,
243 29,88 % e 25,29 % de cobertura de fundo para as densidades 40, 80 e 160 ind.cm⁻²,
244 respectivamente. Para pós-larvas de *A. brasiliiana* a densidade de 160 ind.cm⁻² seria
245 recomendado até alcançar 600 µm de comprimento e 40 ind.cm⁻² até alcançar 1mm,
246 mantendo o limite máximo de crescimento.

247 A taxa de crescimento específico em larvas de *Meretrix meretrix* foi inferior a 2%
248 quando estavam no período de metamorfose e assentamento, independente do densidade
249 de estocagem (Liu et al., 2006). No cultivo de pós-larvas de *A. brasiliiana*, aos 21 dias,
250 observou-se queda na TCE (<2%). Aos 24 dias de cultivo as pós-larvas do tratamento

com menor densidade (40 ind.cm^{-2}) alcançaram 1mm de comprimento, aumentando a taxa de crescimento. O aumento na taxa de crescimento não ocorreu nas outras densidades, onde as larvas ainda não haviam alcançado 1mm de comprimento.

As pós-larvas cultivadas em altas densidades apresentaram o menor crescimento. Essa baixa taxa de crescimento diário afetou o comprimento final, pois aos 28 dias de cultivo as pós-larvas cultivadas na maior densidade apresentaram o menor comprimento.

A sobrevivência final foi menor na densidade 160 ind.cm^{-2} . A densidade de estocagem afetou diretamente na sobrevivência de pós-larvas de *A. brasiliiana*. A taxa de sobrevivência de larvas de bivalves diminui continuamente ao longo do tempo, durante o desenvolvimento das fases D-véliger para pé de véliger, mesmo quando as condições de criação se mantiveram favoráveis (Gouda et al., 2006; Magnesen et al., 2006; Saucedo et al., 2007; Velasco e Barros, 2008). No cultivo de larvas de *Argopecten nucleus* há uma queda na sobrevivência quando as larvas são cultivadas em altas densidades (Velasco e Barros, 2008). Variação na taxa de sobrevivência entre 50 % e 90 % é considerada satisfatória para o cultivo de larvas de moluscos (Utting e Spencer, 1991), o que corrobora com a sobrevivência obtida para as densidades de 40 ind.cm^{-2} e 80 ind.cm^{-2} .

5. Conclusão

Na larvicultura de larvas assentadas de *A. brasiliiana* deve-se realizar o manejo na densidade de estocagem no decorrer do crescimento. Para larvas pé de véliger de *A. brasiliiana* a densidade de $160 \text{ larvas.cm}^{-2}$ seria recomendado até alcançar 600 μm de comprimento e $40 \text{ larvas.cm}^{-2}$ até alcançar 1mm, mantendo o limite máximo de crescimento.

A densidade de estocagem afeta diretamente o crescimento e a sobrevivência de larvas assentadas de *A. brasiliiana*. Em um sistema de cultivo fechado a densidade de 40 larvas.cm⁻² pode ser utilizada mantendo a taxa de crescimento específico em torno de 5% ao dia.

6. Referências

- Epelbaum, A., Pearce, C.M., Yuan, S., Plamondon, N., Gurney-Smith, H., 2011. Effects of stocking density and substratum on the survival, growth, burrowing behaviour and shell morphology of juvenile basket cockle, *Clinocardium nuttallii*: implications for nursery seed production and field out planting. *Aquac. Res.* 42, 975-986.
- Fréchette, M., 2005. A comment on the methodology of stocking experiments. *Aquaculture* 250, 291–299.
- Galley, T.H., Batista, F.M., R. Braithwaite, King, J., Beaumont, A.R., 2010. Optimisation of larval culture of the mussel *Mytilus edulis* (L.). *Aquacult. Int.* 18, 315–325.
- Gouda, R., Kenchington, E., Hatcher, B., Vercaemer, B., 2006. Effects of locally-isolated micro-phytoplankton diets on growth and survival of sea scallop (*Placopecten magellanicus*) larvae. *Aquaculture* 259, 169–180.
- Heasman, M.P., O'Connor, W.A., Frazer, A.W., Languet, Y., O'Connor, S.J., 2002. Alternative means of nursery culture for comercial scallop (*Pecten fumatus* Reeve) spat. *Aquaculture* 213, 323–338.
- Lavander, H.D., Cardoso Jr., L.O., Oliveira, R.L., Silva Neto, S.R., Gálvez, A.O., Peixoto, S.R.M., 2011. Biologia reprodutiva da *Anomalocardia brasiliiana* (Gmelin, 1791) no litoral norte de Pernambuco, Brasil. *Agrária* 6 (2), 344-350.

- 300 Liu, B., Dong, B., Tang, B., Zhang, T., Xiang, J., 2006. Effect of stocking density on
301 growth, settlement and survival of clam larvae, *Meretrix meretrix*. Aquaculture
302 258, 344–349.
- 303 Liu, W., Ma, Y., Hu, S., Miao, G., Li, J., 2002. Rearing Venus Clam seeds, *Cyclina*
304 *sinensis* (Gmelin), on a commercial scale. Aquaculture 211, 109-114.
- 305 Liu, W., Alabi, A.O., Pearce, C.M., 2008. Broodstock conditioning in the basket cockle
306 *Clinocardium nuttallii*. J. Shellfish Res. 27, 399–404.
- 307 Liu, W., Gurney-Smith, H., Beerens, A., Pearce, C.M., 2010. Effects of stocking
308 density, algal density, and temperature on growth and survival of larvae of the
309 basket cockle, *Clinocardium nuttallii*. Aquaculture 299, 99-105.
- 310 Liu, W., Pearce, C.M., Alabi, A.O., Beerens, A., Gurney-Smith, H., 2011. Effects of
311 stocking density, ration, and temperature on growth of early post-settled juveniles
312 of the basket cockle, *Clinocardium nuttallii*. Aquaculture 320, 129–136.
- 313 Loosanoff, V.L., Davis, H.C., 1963. Rearing of bivalve molluscs. In: Russell, F.S. (Ed.),
314 Advances in Marine Biology, vol. 1. Academic Press, London, pp. 1-136.
- 315 Magnesen, T., Bergh, Ø., Christophersen, G., 2006. Yields of great scallops *Pecten*
316 *maximus*, larvae in a commercial flow-through rearing system in Norway. Aquac.
317 Int. 14, 377-394.
- 318 Monti, D., Frenkiel, L., Moueza, M., 1991. Demography and growth of *Anomalocardia*
319 *brasiliiana* (Gmelin) (Bivalvia: Veneridae) in a Mangrove, in Guadeloupe (French
320 West Indies). J. Moll. Stud. 57, (2), 249-257.
- 321 Moueza, M., Gros, O., Frenkiel, L., 1999. Embryonic, larval and postlarval
322 development of the tropical clam, *Anomalocardia brasiliiana* (Bivalvia,
323 Veneridae). J Moll Stud. 65, 73-88.

- 324 Nicolas, L., Robert, R., 2001. The effect of supply on metamorphosis and post-larval
325 development in hatchery-reared *Pecten maximus*. Aquaculture 192, 347-359.
- 326 Nishida, A.K., Nordi, N., Alves, R.R.N., 2004. Abordagem etnoecológica da coleta de
327 moluscos no litoral paraibano. Tropical Oceanography 32 (1), 53-68.
- 328 Oliveira, I., Amorim, A., Lavander, H., Peixoto, S., Galvéz, A. O., 2011. Spatial and
329 temporal distribution of the shellfish *Anomalocardia brasiliiana* (Gmelin, 1791) on
330 Mangue Seco beach, Pernambuco, Brazil. International Journal of Aquatic Science
331 2 (1), 68-79.
- 332 Oliveira, I.B., Silva Neto, S.R., Lima Filho, J.V.M., Peixoto, S.R.M., Gálvez, A.O.,
333 2014. Efeito do período chuvoso na extração do molusco bivalve *Anomalocardia*
334 *brasiliiana* (Gmelin, 1791). Agrária 9 (1), 139-145.
- 335 Pettersen, A. K., Turchini, G.M., Jahangard, S., Ingram, B.A., Sherman, C.D.H., 2010.
336 Effects of diferente dietary microalgae on survival, growth, settlement and fatty
337 acid composition of blue mussel (*Mytilus galloprovincialis*) larvae. Aquaculture
338 309, 115–124.
- 339 Raghavan, G., Gopinathan, C.P., 2008. Effects of diets, stocking density and
340 environmental factors on growth, survival and metamorphosis of clam, *Paphia*
341 *malabarica* (Chamnitz) larvae. Aquac. Res. 39, 928-933.
- 342 Rico-Villa, B., LeCoz, J.R., Mingant, C., Robert, R., 2006. Influence of phytoplankton
343 diet mixtures on microalgae consumption, larval development and settlement of
344 the Pacific oyster *Crassostrea gigas* (Thunberg). Aquaculture 256, 377–388.
- 345 Rupp, G.S., Parson, G.J., 2004. Effects of salinity and temperature on the survival and
346 byssal attachment of the lion’s paw scallop *Nodipecten nodosus* at its southern
347 distribution limit. J. Exp. Mar. Biol. Ecol. 309, 173-198.

- 348 Saucedo, P.E., Ormart-Castro, P., Osuna-García, P., 2007. Towards development of
349 large-scale hatchery cultivation of larvae and spat of the pearl oyster *Pinctada*
350 *mazatlanica* in Mexico. *Aquaculture* 273, 478-486.
- 351 Silva-Cavalcanti, J.S., Costa, M.F., 2009. Fisheries in protected and non-protected
352 areas: is it different? The case of *Anomalocardia brasiliiana* at tropical estuaries of
353 Northeast Brazil. *J. Coastal Res.* SI 56 (Proceedings of the 10th International
354 Coastal Symposium), 1454 – 1458.
- 355 Sprung, M., 1984a. Physiological energetics of mussel larvae (*Mytilus edulis*). I. Shell
356 growth and biomass. *Mar. Ecol. Prog. Ser.* 17, 283–293.
- 357 Sprung, M., 1984b. Physiological energetics of mussel larvae (*Mytilus edulis*). II. Food
358 up take. *Mar. Ecol. Prog. Ser.* 17, 295–305.
- 359 Utting, S.D., Spencer, B.E., 1991. The hatchery culture of bivalve mollusk larvae and
360 juveniles. *Lab. Leaflet, MAFF Fish. Res., Lowestoft*, (68), 31 pp.
- 361 Velasco, L.A., Barros, J., 2008. Experimental larval culture of the Caribbean scallops
362 *Argopecten nucleus* and *Nodipecten nodosus*. *Aquac. Res.* 39, 603–618.
- 363
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Tabela 1. Concentração algal fornecida às larvas de *A. brasiliiana* até atingir o estágio juvenil.

Etapas	Dias	Fases	Concentração Algal
1	1 - 7	D véliger / Umbonada / véliger	30.000 cél.mL ⁻¹
2	8 - 12	Pé de véliger	40.000 cél.mL ⁻¹
3	13 - 45	Pé de véliger / Juvenil	50.000 cél.mL ⁻¹

Tabela 2. Taxa de crescimento específico de larvas pós-assentamento de *A. brasiliiana* em três diferentes densidades de estocagem para cada período cultivado (dias).

TCE (%.dia ⁻¹)					
Densidade de estocagem (Larvas.cm ⁻²)	Período do cultivo (dias)				
	0-7	7-14	14-21	21-28	0-28
40	8,21±0,33 ^a	6,00±0,32 ^a	0,37±0,32 ^b	5,02±0,32 ^a	4,98±0,08 ^a
80	8,44±0,27 ^a	3,91±0,30 ^b	1,47±0,26 ^a	2,27±0,30 ^b	4,08±0,07 ^b
160	8,52±0,23 ^a	2,73±0,23 ^c	1,19±0,34 ^{ab}	0,82±0,29 ^c	3,37±0,07 ^c

Médias com caracteres diferentes diferem significativamente (p<0,05).

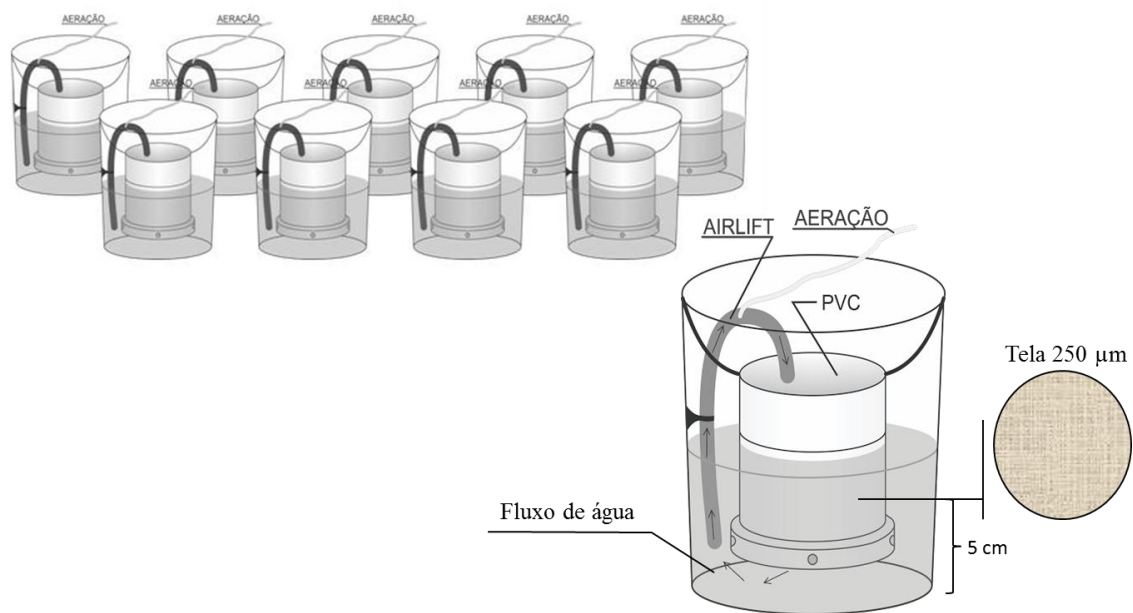


Figura 1. Desenho esquemático das unidades experimentais utilizadas para avaliação do efeito da densidade de estocagem no crescimento e sobrevivência de larvas pé de véliger de *A. brasiliana*.

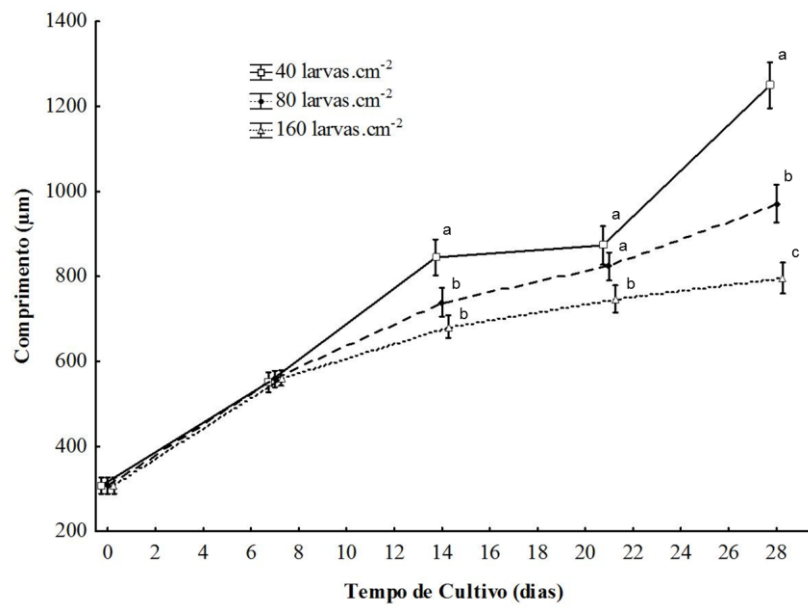


Figura 2. Comprimento médio ($\pm$ EP) de larvas pé de véliger de *A. brasiliana* para cada período do cultivo (7, 14, 21 e 28 dias) em três diferentes densidades de estocagem. Médias com letras sobrescritas diferentes diferem significativamente ($p < 0,05$).

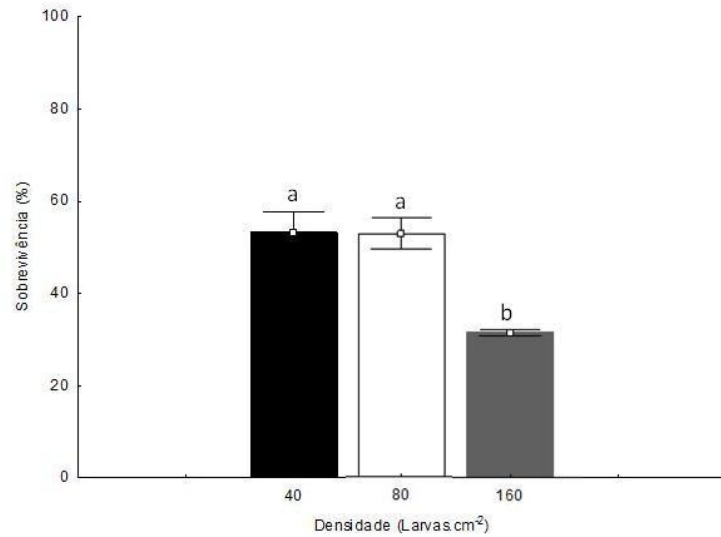


Figura 3. Sobrevivência larval de *Anomalocardia brasiliiana* cultivada por 28 dias em diferentes densidades de estocagem. Médias com caracteres diferentes diferem significativamente ($p < 0,05$).

6. CONSIDERAÇÕES FINAIS

A larvicultura do molusco bivalve *Anomalocardia brasiliiana* pode ser realizada utilizando diversos tipos de dietas microalagais, porém a microalga *I. galbana* quando fornecida isoladamente não apresentou crescimento e sobrevivência larval satisfatório, devendo ser utilizada sempre combinada a uma outra espécie, principalmente, diatomáceas.

No cultivo de pós-larvas do molusco bivalve *A. brasiliiana* deve-se considerar o manejo na densidade de estocagem no decorrer do crescimento das pós-larvas. No início da metamorfose para pé-de-véliger pode-se utilizar a densidade de 160 ind.cm² até que estes alcancem 600µm de comprimento (eixo dorso ventral), posteriormente reduzir a densidade de estocagem para 40 ind.cm² até 1mm de comprimento.

A fim de melhorar os resultados obtidos neste estudo, novos trabalhos deverão ser realizados com objetivo de determinar o melhor sistema de cultivo, aberto ou fechado com recirculação, diferentes condições de temperatura e salinidade, além do uso de probióticos na larvicultura de *A. brasiliiana*.

ANEXOS

Normas da Revista Latin American Journal of Aquatic Research

Instructions for Authors

General Publishing Instructions for the Authors

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- Results
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- Conclusions (optional)
- Acknowledgements

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- Coelho, V., R.A. Cooper & S. Rodrigues. 2000. Burrow morphology and behaviour of the mud shrimp *Upogebia omissa* (Decapoda, Thalassinidea, Upogebiidae). Mar. Ecol. Progr. Ser., 200: 229-240.

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- Thurman, H. & A. Trujillo. 2002. Essentials of oceanography. Prentice Hall, New Jersey, 524 pp.

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- Brummet, R.E. & B.A. Costa-Pierce. 2002. Village-based aquaculture ecosystems as a model for sustainable aquaculture development in Sub-Saharan Africa. In: B. Costa-Pierce (ed.). Ecological aquaculture: evolution of the blue revolution. Blackwell Science, Oxford, pp. 145-160.

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- Retamal, M.A. 2000. (CD-ROM). Decápodos de Chile. ETI-Universidad de Concepción. Springer-Verlag, Berlin.

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A review article is a scientific paper which provides a synthesis of current state information on specific research topic. The review consists of an abstract (maximum 200 words) and keywords written in English and Spanish, plus a continuous free-style section. The manuscript should not exceed 20 pages including the figures and tables.

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Review Articles can cover either narrow disciplinary subjects or broad issues requiring interdisciplinary discussion. They should provide objective critical evaluation of a defined subject. Reviews should not consist solely of a summary of published data. Evaluation of the quality of existing data, the status of knowledge, and the research required to advance knowledge of the subject are essential.

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The Letters to the Editor section is intended to provide a forum for discussion of aquacultural science emanating from material published in the journal.

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D.M. Gatlin

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- 1) determination of dietary and metabolic requirements for various nutrients by representative aquatic species. Studies may include environmental/stress effects on animal's physiological responses and requirements at different developmental stages;

- 2) evaluation of novel or established feedstuffs as well as feed processing and manufacturing procedures with digestibility and growth trials. Such studies should provide comprehensive specifications of the process or evaluated ingredients including nutrients, potential anti-nutrients, and contaminants;
- 3) comparison of nutrient bioavailability from various ingredients or product forms as well as metabolic kinetics of nutrients, food borne anti-nutrients or toxins;
- 4) identification of key components in natural diets that influence attractability, palatability, metabolism, growth reproduction and/or immunity of cultured organisms;
- 5) optimization of diet formulations and feeding practices;
- 6) characterization of the actions of hormones, cytokines and/or components in intracellular signaling pathway(s) that influence nutrient and/or energy utilization;
- 7) evaluation of diet supplementation strategies to influence animal performance, metabolism, health and/or flesh quality.

Manuscripts concerning other areas of nutrition using novel or advanced methods are also welcome. Please note that in regard to various diet additives such as probiotics, prebiotics, herbal extracts, etc., a very large number of papers have already been published. Therefore, Aquaculture will not continue to accept manuscripts that present initial and preliminary investigations of such additives. Manuscripts addressing these and other feed additives will be accepted for review only if they are of the highest scientific quality and they represent a significant advance in our knowledge of the mechanisms involved in their metabolism. Manuscripts may also be considered if they present clinical efficacy data generated in large-scale trials and economic cost-benefit analysis of these applications.

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B.Costa-Pierce

AQUACULTURE PRODUCTION SCIENCE (PS) is one of 5 sections of the international journal AQUACULTURE dedicated to research on improvements and innovations in aquatic food production.

This section supports worldwide dissemination of the results of innovative, globally important, scientific research on production methods for aquatic foods from fish, crustaceans, mollusks, amphibians, and all types of aquatic plants. Contributions are encouraged in the following areas: 1) Improvement of production systems that results in greater efficiencies of resource usage and sustainability of aquaculture; 2) Effective applications of technologies and methods of aquaculture production for improved stocking regimes; 3) The use of new species and species assemblages; and, 4) Investigations to minimize aquaculture wastes and improve water quality, including technologies for nutrient recycling in aquaculture ecosystems, and potential synergy of aquaculture and other food production systems using methods such as polyculture and integrated aquaculture. Aspects of seafood processing and technology will not be considered in this section although aquaculture techniques that may influence the

nutritional value of aquatic food products may be considered in the Nutrition Section.

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Fish: A. P. (Tony) Farrell

Invertebrates: J. Benzie

The Physiology Section welcomes high quality papers that present either novel research data or original reviews. The content must be relevant to solving aquaculture problems on all aspects of the physiology of cultured aquatic animals and plants.

Submitted manuscripts must have a valid hypothesis or objective, clearly state the relevance to aquaculture, have proper experimental design with appropriate controls and utilize appropriate statistical analysis. Mention of trade names is limited to the main text.

Relevant physiological topics include, but are not limited to:

- Reproductive and endocrine physiology, including control of development and sex differentiation, induced ovulation and spermiation, gamete quality, storage and cryopreservation, physiology of gynogenetic, and triploid and transgenic organisms
- Cardiorespiratory, muscle and exercise physiology
- Osmoregulatory physiology
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- Larval physiology and ontogeny, including metamorphosis, smolting and molting
- Performance under variable culture conditions, including temperature, water quality, rearing density, and stress and disease physiology
- Physiology of harvest and handling techniques

Genetics:

G. Hulata

The Genetics Section welcomes high-quality research papers presenting novel data, as well as critical reviews, on various aspects of selective breeding, genetics and genomics. Submitted manuscripts must have a valid hypothesis or objective, clearly state the relevance to aquaculture, have proper experimental design with appropriate sample size and controls and utilize appropriate statistical analysis.

Relevant genetics topics include, but are not limited to:

- Breeding programs using classic selection procedures, markers or combining marker assisted selection with classic selection
- Applications of crossbreeding and interspecific hybridization
- Evaluation of commercially important phenotypes among cultured strains, populations or stocks
- Applications of biotechnology and genetic manipulation methods
- Development of linkage maps, identification of QTL or association of commercially important traits with specific gene(s). Where appropriate, linkage maps should include co-dominant markers, such as microsatellite DNA and SNP

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Aquaculture will NOT accept manuscripts dealing with the application of well-described techniques to yet another species, unless the application solves a specific biological problem important to aquaculture production; or manuscripts dealing with gene cloning, characterizing of microsatellites, species identification using molecular markers, EST papers with small collections, or mapping papers with a small number of markers, unless the papers also deal with solving a biological problem that is relevant to aquaculture production.

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Sustainability and Society:

D.C. Little

The Sustainability and Society section of the journal Aquaculture invites articles at the interface of natural and social sciences that address the broader roles of aquaculture in global food security and trade.

Aims and scope of the Sustainability and Society section are the: global dissemination of interdisciplinary knowledge regarding the management of aquatic resources and resulting impacts on people. Interconnections with other sectors of food production; resource management and implications for societal impact. Going beyond a narrow techno-centric focus, towards more holistic analyses of aquaculture within well-defined contexts. Enquiry based on understanding trajectories of change amid the global challenges of climate change and food security. Mixed methods and approaches that incorporate and integrate both social and natural sciences. Relevance for the diverse range of policy makers, practitioners and other stakeholders involved. Articles that take a value chain approach, rather than being wholly production orientated, are encouraged.

Disease

B. Austin

The Disease sections welcomes critical reviews and high quality articles containing novel data on all aspects concerning diseases of farmed aquatic species. The aims of the section are: description of new and emerging diseases including characterization of the causal agent(s), development in the understanding of fish pathogens for example including new methods of growth where this has been a problem for fastidious organisms, pathogenicity and epizootiology, developments in the diagnosis of disease going beyond the use of standard well used methods, and methods of disease control, notably new developments in vaccines, immunostimulants, dietary supplements, medicinal plant products, probiotics, prebiotics and genetically-disease resistant stock. Relevance to aquaculture must be demonstrated. Articles, which adapt well known methods without further refinement of those methods, are unlikely to be accepted.



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